BD™ Cytometric Bead Array (CBA)

Mouse Th1/Th2 Cytokine Kit

Cat. No. 551287

BD

Kit Contents

80 Tests (50 Samples and 2 standard curves)

(Store the following items at 4°C)

A1 Mouse IL-2 Capture Beads: 1 vial, 0.8 ml

A2 Mouse IL-4 Capture Beads: 1 vial, 0.8 ml

A3 Mouse IL-5 Capture Beads: 1 vial, 0.8 ml

A4 Mouse IFN-γ Capture Beads: 1 vial, 0.8 ml

A5 Mouse TNF Capture Beads: 1 vial, 0.8 ml

B Mouse Th1/Th2 PE Detection Reagent: 1 vial, 4 ml

C Mouse Th1/Th2 Cytokine Standards: 2 vials, 0.2 ml lyophilized

D Cytometer Setup Beads: 1 vial, 1.5 ml

E1 PE Positive Control Detector: 1 vial, 0.5 ml

E2 FITC Positive Control Detector: 1 vial, 0.5 ml

F Wash Buffer: 1 bottle, 130 ml

G Assay Diluent: 1 bottle, 30 ml

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Introduction

Flow cytometry is an analysis tool that allows for the discrimination of different particles on the basis of size and color. Multiplexing is the simultaneous assay of many analytes in a single sample. The BD™ Cytometric Bead Array (CBA) employs a series of particles with discrete fluorescence intensities to simultaneously detect multiple soluble analytes. The BD CBA is combined with flow cytometry to create a powerful multiplexed assay.

The BD CBA system uses the sensitivity of amplified fluorescence detection by flow cytometry to measure soluble analytes in a particle-based immunoassay. Each bead in a BD CBA Kit provides a capture surface for a specific protein and is analogous to an individually coated well in an ELISA plate. The BD CBA capture bead mixture is in suspension to allow for the detection of multiple analytes in a small sample volume. The combined advantages of the broad dynamic range of fluorescent detection via flow cytometry and the efficient capturing of analytes via suspended particles enable the BD CBA to use fewer sample dilutions and to obtain the value of an unknown in substantially less time (compared to conventional ELISA).

The BD Mouse Th1/Th2 Cytokine CBA Kit can be used to measure Interleukin-2 (IL-2), Interleukin-4 (IL-4), Interleukin-5 (IL-5), Interferon-γ (IFN-γ), and Tumor Necrosis Factor (TNF) protein levels in a single sample. The kit performance has been optimized for analysis of physiologically relevant concentrations (pg/ml levels) of specific cytokine proteins in tissue culture supernatants and serum samples.

Principle of the Test

Five bead populations with distinct fluorescence intensities have been coated with capture antibodies specific for IL-2, IL-4, IL-5, IFN-γ, and TNF proteins. The five bead populations are mixed together to form the BD CBA, which is resolved in a red channel (ie, FL3 or FL4) of a flow cytometer.

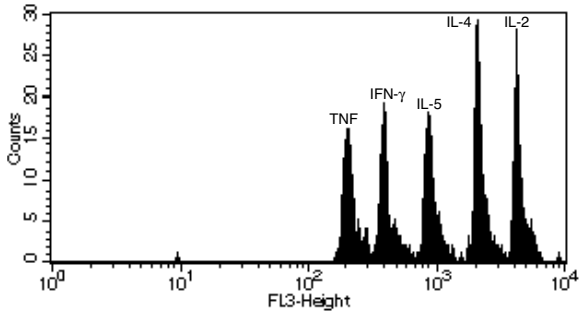


Figure 1

The cytokine capture beads are mixed with the PE-conjugated detection antibodies and then incubated with recombinant standards or test samples to form sandwich complexes. Following acquisition of sample data using the flow cytometer, the sample results are generated in graphical and tabular format using the BD CBA Analysis Software or FCAP Array™ Software. The kit provides sufficient reagents for the quantitative analysis of 50 test samples and the generation of two standard curve sets.

Advantages

The BD CBA provides several advantages when compared with conventional ELISA methodology:

- The required sample volume is approximately one-fifth the quantity necessary for conventional ELISA assays due to the detection of five analytes in a single sample.
- A single set of diluted standards is used to generate a standard curve for each analyte.
- A BD CBA experiment takes less time than a single ELISA and provides results that would normally require five conventional ELISAs

Limitations

The theoretical limit of detection of the BD CBA Mouse Th1/Th2 Cytokine kit is comparable to conventional ELISA, but due to the complexity and kinetics of this multi-analyte assay, actual limit of detection on a given experiment may vary slightly (see ***Theoretical Limit of Detection*** and ***Precision*** page 21 and 25 respectively).

The BD CBA is not recommended for use on stream-in-air instruments where signal intensities may be reduced, adversely effecting assay sensitivity. Stream-in-air instruments include the BD FACStar™ Plus and BD FACSVantage™ (BD Immunocytometry Systems, San Jose, CA) flow cytometers.

Serum spike recoveries for IL-4 and TNF are lower than for the other cytokines in this assay. This variation is due to assay conditions and serum proteins and may affect quantitation of these proteins in serum samples.

Quantitative results or protein levels for the same sample or recombinant protein run in ELISA and BD CBA assays may differ. A spike recovery assay can be performed using an ELISA standard followed by BD CBA analysis to assess possible differences in quantitation.

This kit is designed to be used as an integral unit. Do not mix components from different batches or kits.

Reagents Provided

Bead Reagents

Mouse Cytokine Capture Beads (A1 – A5): The specific capture beads, having discrete fluorescence intensity characteristics, are distributed from brightest to dimmest as follows:

Bead	Specificity
(Brightest) A1	IL-2
A2	IL-4
A3	IL-5
A4	IFN-γ
(Dimmest) A5	TNF

A single 80-test vial of each specific capture bead (A1 – A5) is included in this kit. Store at 4°C. Do not freeze.

Note: The antibody-conjugated beads will settle out of suspension over time. It is necessary to vortex the vial vigorously for 3 - 5 seconds before taking a bead suspension aliquot.

Cytometer Setup Beads (D): A single 30-test vial of setup beads for setting the initial instrument PMT voltages and compensation settings is sufficient for 10 instrument setup procedures. The Cytometer Setup Beads are formulated for use at 50 µl/test.

Antibody and Standard Reagents

Mouse Th1/Th2 PE Detection Reagent (B): An 80-test vial of PE-conjugated anti-mouse IL-2, IL-4, IL-5, IFN-γ, and TNF antibodies that is formulated for use at 50 µl/test. Store at 4°C. Do not freeze.

PE Positive Control Detector (E1): A 10-test vial of PE-conjugated antibody control that is formulated for use at 50 µl/test. This reagent is used with the Cytometer Setup Beads to set the initial instrument compensation settings. Store at 4°C. Do not freeze.

FITC Positive Control Detector (E2): A 10-test vial of FITC-conjugated antibody control that is formulated for use at 50 µl/test. This reagent is used with the Cytometer Setup Beads to set the initial instrument compensation settings. Store at 4°C. Do not freeze.

Mouse Th1/Th2 Cytokine Standards (C): Two vials containing lyophilized recombinant mouse cytokine proteins. Each vial should be reconstituted in 2.0 ml of Assay Diluent to prepare the top standard. Store at 4°C.

Buffer Reagents

Assay Diluent (G): A single 30 ml bottle of a buffered protein* solution (1×) used to reconstitute and dilute the Mouse Th1/Th2 Cytokine Standards and to dilute test samples. Store at 4°C.

Wash Buffer (F): A single 130 ml bottle of phosphate buffered saline (PBS) solution (1×), containing protein* and detergent, used for wash steps and to resuspend the washed beads for analysis. Store at 4°C.

Warnings and Precautions

Hazardous Ingredients:

Sodium Azide:

Components A1-A5, B, D, E1-E2, F, and G contain 0.09% sodium azide. Sodium azide yields a highly toxic hydrazoic acid under acidic conditions. Avoid exposure to skin and eyes, ingestion, and contact with heat, acids, and metals. Wash exposed skin with soap and water. Flush eyes with water. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.

** Source of all serum proteins is from USDA inspected abattoirs located in the United States.*

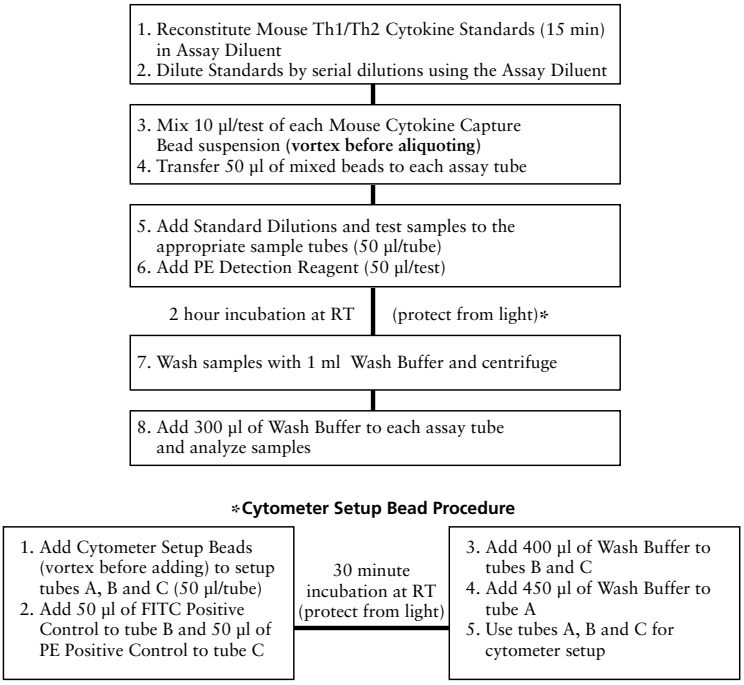
Materials Required but Not Provided

In addition to the reagents provided in the BD CBA Mouse Th1/Th2 Cytokine Kit, the following items are also required:

- A flow cytometer equipped with a 488 nm laser capable of detecting and distinguishing fluorescence emissions at 576 and 670 nm (eg, BD FACScan™ or BD FACSCalibur™ instruments) and BD CellQuest™ software.
- 12 × 75 mm sample acquisition tubes for a flow cytometer (eg, BD Falcon™ Cat. No. 352008).
- BD CBA Software, or FCAP Array software (Cat. No. 641488).

  Note: The BD CBA Software is no longer available for purchase but is still supported on existing compatible systems.
- BD Calibrite™ 3 Beads, (Cat. No. 340486).

Overview: BD CBA Mouse Th1/Th2 Cytokine Assay Procedure



Preparation of Mouse Th1/Th2 Cytokine Standards

The Mouse Th1/Th2 Cytokine Standards are lyophilized and should be reconstituted and serially diluted before mixing with the Capture Beads and the PE Detection Reagent.

1. Open one vial of lyophilized Mouse Th1/Th2 Standards. Transfer the standard spheres to a polypropylene tube (eg, 15 ml Conical Tube, BD Falcon Cat. No. 352097). Label tube "Top Standard".
2. Reconstitute the standards with 2.0 ml of Assay Diluent. Allow the reconstituted standard to equilibrate for at least 15 minutes before making dilutions. **Mix reconstituted protein by pipette only. Do not vortex or mix vigorously.**
3. Label 12 × 75 mm tubes (BD Falcon Cat. No. 352008) and arrange them in the following order: 1:2, 1:4, 1:8, 1:16, 1:32, 1:64, 1:128, and 1:256.
4. Pipette 300 µl of Assay Diluent to each of the remaining tubes.
5. Perform a serial dilution by transferring 300 µl from the Top Standard to the 1:2 dilution tube and mix thoroughly. Continue making serial dilutions by transferring 300 µl from the 1:2 tube to the 1:4 tube and so on to the 1:256 tube and mix thoroughly (see ***Figure 2***). **Mix by pipette only, do not vortex.** Prepare one tube containing Assay Diluent to serve as the 0 pg/ml negative control.

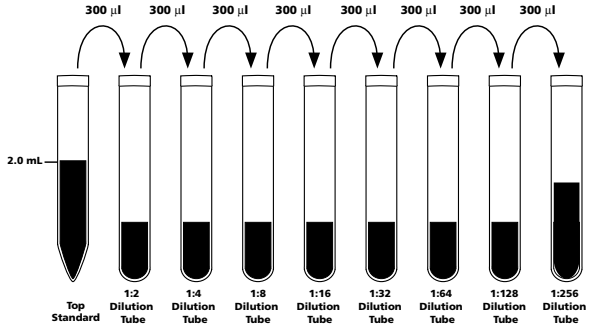


Figure 2. Preparation of Mouse Th1/Th2 Cytokine Standard Dilutions

The approximate concentration (pg/ml) of recombinant protein in each dilution tube is shown in ***Table 1.***

Table 1. Mouse Th1/Th2 Cytokine Standard concentrations after dilution

Protein (pg/ml)	Top Standard	1:2 Dilution Tube	1:4 Dilution Tube	1:8 Dilution Tube	1:16 Dilution Tube	1:32 Dilution Tube	1:64 Dilution Tube	1:128 Dilution Tube	1:256 Dilution Tube
Mouse IL-2	5000	2500	1250	625	312.5	156	80	40	20
Mouse IL-4	5000	2500	1250	625	312.5	156	80	40	20
Mouse IL-5	5000	2500	1250	625	312.5	156	80	40	20
Mouse IFN-γ	5000	2500	1250	625	312.5	156	80	40	20
Mouse TNF	5000	2500	1250	625	312.5	156	80	40	20

Preparation of Mixed Mouse Th1/Th2 Cytokine Capture Beads

The Capture Beads are bottled individually and it is necessary to pool the bead reagents (A1 – A5) immediately before mixing them together with the standards, samples and PE Detection reagent.

1. Determine the number of assay tubes (including standards and controls) that are required for the experiment (eg, 8 unknowns, 9 cytokine standard dilutions and 1 negative control = 18 assay tubes).
2. Vigorously vortex each Capture Bead suspension for a few seconds before mixing.
3. Add a 10 µl aliquot of each Capture Bead, for each assay tube to be analyzed, into a single tube labeled "mixed Capture Beads" (eg, 10 µl of IL-2 Capture Beads × 18 assay tubes = 180 µl of IL-2 Capture Beads required).

   Note: Extra tests of Capture Beads should be mixed to ensure that the necessary number of tests will be recovered from the mixed Capture Beads tube. (eg, add an additional 2 – 3 assay tubes to the number determined in step 1 above before calculating the amount to add to the mixed Capture Beads tube in step 3).

4. Vortex the Bead mixture thoroughly.

The mixed Capture Beads are now ready to be transferred to the assay tubes (50 µl of mixed Capture Beads/tube) as described in ***Mouse Th1/Th2 Cytokine CBA Assay Procedure***, page 12.

Note: Discard excess mixed Capture Beads. Do not store after mixing.

Preparation of Test Samples

The standard curve for each cytokine covers a defined set of concentrations from 20 – 5000 pg/ml. It may be necessary to dilute test samples to ensure that their mean fluorescence values fall within the limits or range of the generated cytokine standard curve. For best results, samples that are known or assumed to contain high levels of a given cytokine should be diluted as described below.

1. Dilute test sample by the desired dilution factor (ie, 1:2, 1:10 or 1:100) using the appropriate volume of Assay Diluent.
2. Mix sample dilutions thoroughly before transferring samples to the appropriate assay tubes containing mixed Capture Beads and PE Detection Reagent.

BD CBA Mouse Th1/Th2 Cytokine Assay Procedure

Following the preparation and dilution of the standards and mixing of the capture beads, transfer these reagents and test samples to the appropriate assay tubes for incubation and analysis. **In order to calibrate the flow cytometer and quantitate test samples, it is necessary to run the Cytokine Standards and the Cytometer Setup controls in each experiment.** See Table 2 for a detailed description of the reagents added to the Cytokine Standard control assay tubes. The Cytometer Setup procedure is described in ***Cytometer Setup, Data Acquisition and Analysis***, page 13.

1. Add 50 µl of the mixed Capture Beads to the appropriate assay tubes. Vortex the mixed Capture Beads before adding to the assay tubes.
2. Add 50 µl of the Mouse Th1/Th2 Cytokine Standard dilutions to the control assay tubes.
3. Add 50 µl of each test sample to the test assay tubes.
4. Add 50 µl of the Mouse Th1/Th2 PE Detection Reagent to the assay tubes.
5. Incubate the assay tubes for 2 hours at RT and protect from direct exposure to light.
6. Add 1 ml of Wash Buffer to each assay tube and centrifuge at 200 × *g* for 5 minutes.
7. Carefully aspirate and discard the supernatant from each assay tube.
8. Add 300 µl of Wash Buffer to each assay tube to resuspend the bead pellet.
9. Begin analyzing samples on a flow cytometer. **Vortex each sample for 3 - 5 seconds immediately before analyzing on the flow cytometer.**

   Note: It is necessary to analyze CBA samples on the day of the experiment. Prolonged storage of samples, once the assay is complete, can lead to increased background and reduced sensitivity.

Table 2. Essential control assay tubes

Tube No.	Reagents (All reagent volumes are 50 µl)
1 (Negative Control 0 pg/ml Standards)	mixed Capture Beads, Assay Diluent, PE Detection Reagent
2 (20 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:256 Dilution, PE Detection Reagent
3 (40 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:128 Dilution, PE Detection Reagent
4 (80 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:64 Dilution, PE Detection Reagent
5 (156 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:32 Dilution, PE Detection Reagent
6 (312 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:16 Dilution, PE Detection Reagent
7 (625 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:8 Dilution, PE Detection Reagent
8 (1250 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:4 Dilution, PE Detection Reagent
9 (2500 pg/ml Standards)	mixed Capture Beads, Cytokine Standards 1:2 Dilution, PE Detection Reagent
10 (5000 pg/ml Standards)	mixed Capture Beads, Cytokine Standards "Top Standard", PE Detection Reagent

Cytometer Setup, Data Acquisition and Analysis

The Cytometer setup information in this section is for the BD FACScan and BD FACSCalibur flow cytometers. The BD FACSComp software is useful for setting up the flow cytometer. BD CellQuest software is required for analyzing samples and formatting data for subsequent analysis using the BD CBA Software or FCAP Array Software.

Additional setup protocols for the BD FACSCalibur flow cytometer (dual laser), BD FACSArray™ bioanalyzer, and other BD FACS brand flow cytometers can be found at **bdbiosciences.com/cbasetup**.

Preparation of Cytometer Setup Beads

1. Add 50 µl of Cytometer Setup Beads to three cytometer setup tubes labeled A, B and C.
2. Add 50 µl of FITC Positive Control Detector to tube B.
3. Add 50 µl of PE Positive Control Detector to tube C.
4. Incubate tubes A, B and C for 30 minutes at room temperature and protect from direct exposure to light.
5. Add 450 µl of Wash Buffer to tube A and 400 µl of Wash Buffer to tubes B and C.
6. Proceed to next section.

Instrument Setup with BD FACSComp Software and BD Calibrite Beads

1. Perform instrument start up.
2. Perform flow check.
3. Prepare tubes of BD Calibrite beads and open BD FACSComp software.
4. Launch BD FACSComp software
5. Run BD FACSComp software in Lyse/No Wash mode.
6. Proceed to next section.

Note: For detailed information on using BD FACSComp with BD Calibrite beads to set up the flow cytometer, refer to the *BD FACSComp Software User's Guide* and the *BD Calibrite Beads* Package Insert. Version 4.2 contains a BD CBA preference setting to automatically save a BD CBA calibration file at the successful completion of any Lyse/No Wash assay. The BD CBA calibration file provides the optimization for FSC, SSC, and threshold settings as described in ***Instrument Setup with the Cytometer Setup Beads***, Steps 3 – 5. Optimization of the fluorescence parameter settings is still required (ie, PMT and compensation settings, see ***Instrument Setup with the Cytometer Setup Beads***, Step 6).

Instrument Setup with the Cytometer Setup Beads

1. Launch BD CellQuest Software and open the BD CBA Instrument Setup template.

   Note: The BD CBA Instrument Setup template can be found on the BD FACStation CD for Macintosh computers in the BD CBA folder. This file may also be downloaded via the internet from: **bdbiosciences.com/cbatemplates**

2. Set the instrument to Acquisition mode.

   Note: The data will be evaluated in five parameters (FSC, SSC, FL1, FL2 and FL3). Turn off additional detectors.

3. Set SSC (side light scatter) and FSC (forward light scatter) to Log mode.
4. Decrease the SSC PMT voltage by 100 from what BD FACSComp set.
5. Set the Threshold to FSC at 650.
6. In setup mode, run Cytometer Setup Beads tube A. Follow the setup instructions on pages 15 – 17.

   Note: Pause and restart acquisition frequently during the instrument setup procedure in order to reset detected values after settings adjustments.

Adjust gate R1 so that the singlet bead population is located in gate R1 (***Figure 3a***).

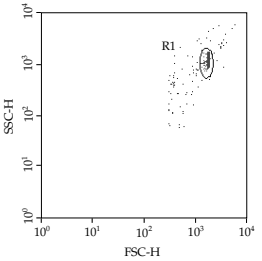


Figure 3a

Adjust the FL3 PMT so that the median of the top FL3 bead population's intensity is around 5000 (***Figure 3b***). Adjust gate R3 as necessary so that the dim FL3 bead population is located in gate R3 (***Figure 3b***). Do not adjust the R2 gate.

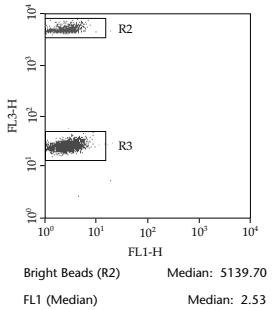


Figure 3b

Adjust the FL1 PMT so that the median of FL1 is approximately 2.0 – 2.5 (***Figure 3b***).

Adjust the FL2 PMT value so that the median of FL2 is approximately 2.0 – 2.5 (***Figure 3c***)**.**

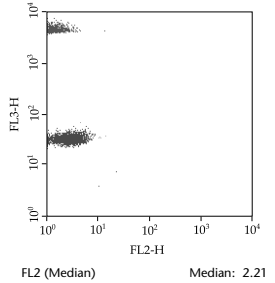


Figure 3c

Run Cytometer Setup Beads tube B to adjust the compensation settings for FL2 – %FL1.

Adjust gate R5 as necessary so that the FL1 bright bead population is located in gate R5 (***Figure 3d***). Using the FL2 – %FL1 control, adjust the median of R5 to equal the median of R4 (***Figure 3d***).

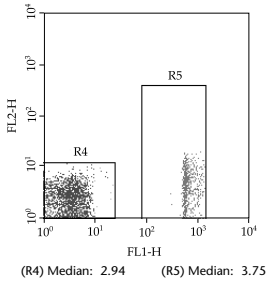


Figure 3d

Run Cytometer Setup Beads tube C to adjust the compensation settings for FL1 – %FL2 and FL3 – %FL2.

Adjust gate R7 so that the FL2 bright bead population is located in gate R7 (***Figure 3e***). Using the FL1 – %FL2 control, adjust the median of R7 to equal the median of R6 (***Figure 3e***).

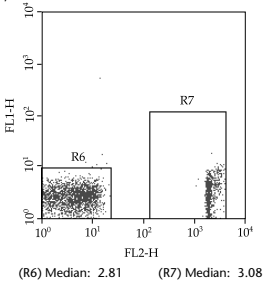


Figure 3e

Adjust gate R9 so that the FL2 bright bead population is located in gate R9 (***Figure 3f***). Using the FL3 – %FL2 control, adjust the median of R9 to equal the median of R8 (***Figure 3f***).

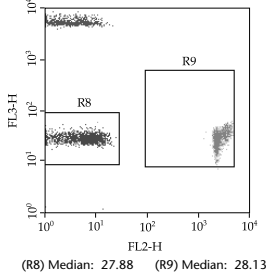


Figure 3f

Set the FL2 – %FL3 to 0.1 if necessary. Save and print the optimized instrument settings.

Data Acquisition

1. Open the acquisition template.

   Note: The acquisition template may be downloaded via the internet from: **bdbiosciences.com/cbatemplates**

2. Set acquisition mode and retrieve the optimized instrument settings from ***Instrument Setup with the Cytometer Setup Beads***, page 14.

3. In the Acquisition and Storage window, set the resolution to 1024.

4. Set number of events to be counted at 1500 of R1 gated events. (This will ensure that the sample file contains approximately 300 events per Capture Bead).

5. Set number of events to be collected to "all events". Saving all events collected will ensure that no true bead events are lost due to incorrect gating.

6. In setup mode, run tube no. 1 and using the FSC vs. SSC dot plot, place the R1 region gate around the singlet bead population (see ***Figure 3a***).

7. Samples are now ready to be acquired.

8. Begin sample acquisition with the flow rate set at HIGH.

   Note: Run the negative control tube (0 pg/ml standards) before any of the recombinant standard tubes. Run the control assay tubes before any unknown test assay tubes. Run the tubes in the order listed in Table 2, page 12.

   File names must be alphanumeric (ie, contain at least one letter).

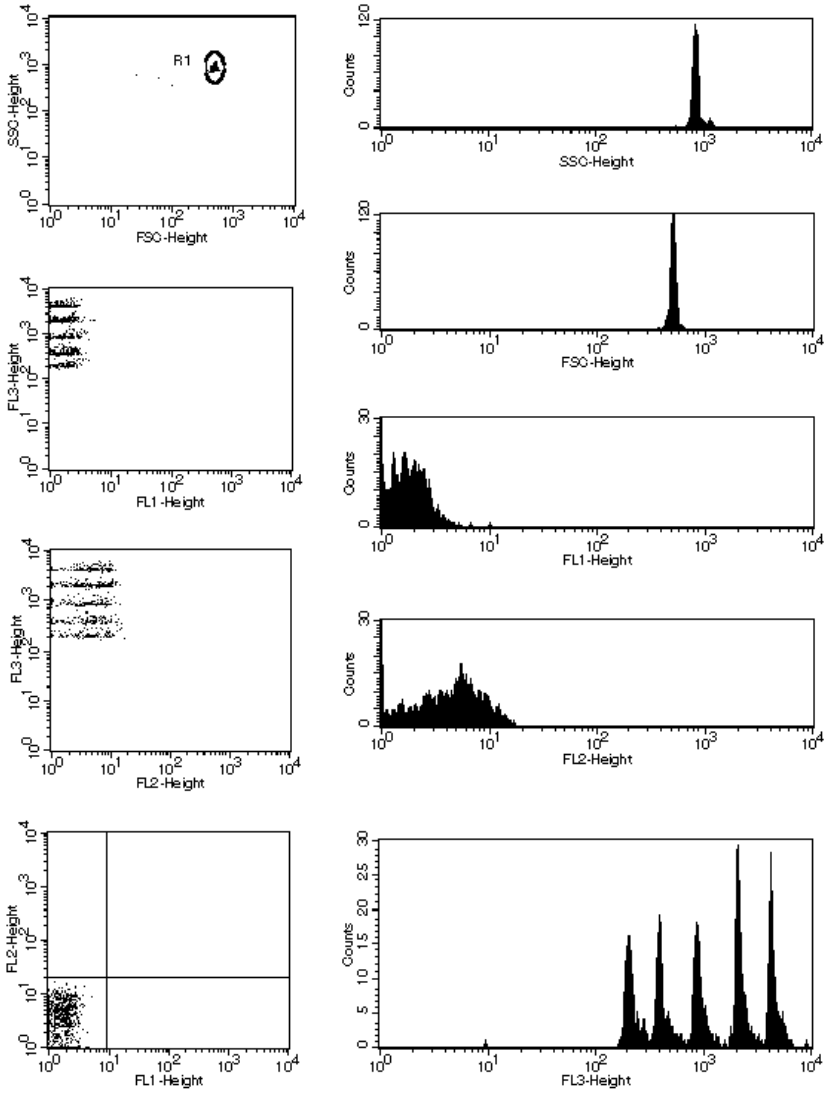


Figure 4. Acquisition Template Example

Analysis of Sample Data

The analysis of BD CBA data can be accomplished using BD CBA Software or FCAP Array software. For BD CBA Software, please refer to the User's Guide for instructions. For FCAP Array software, please visit the following link for instructions: **bdbiosciences.com/docs/kitanalysis**

Typical Data

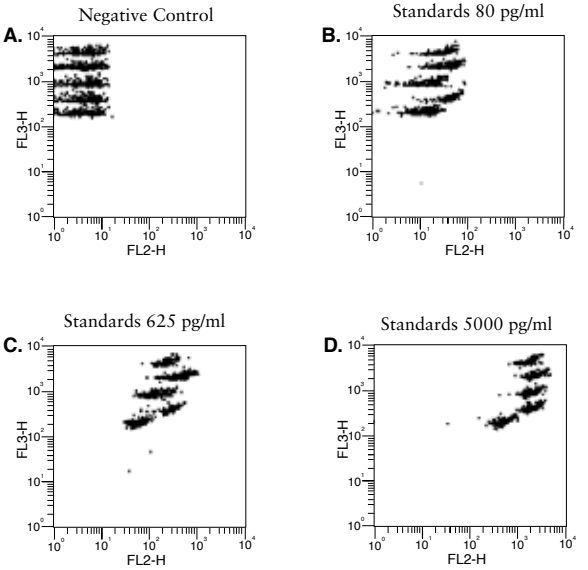


Figure 5. BD CellQuest Data Examples for Standards and Detectors Alone

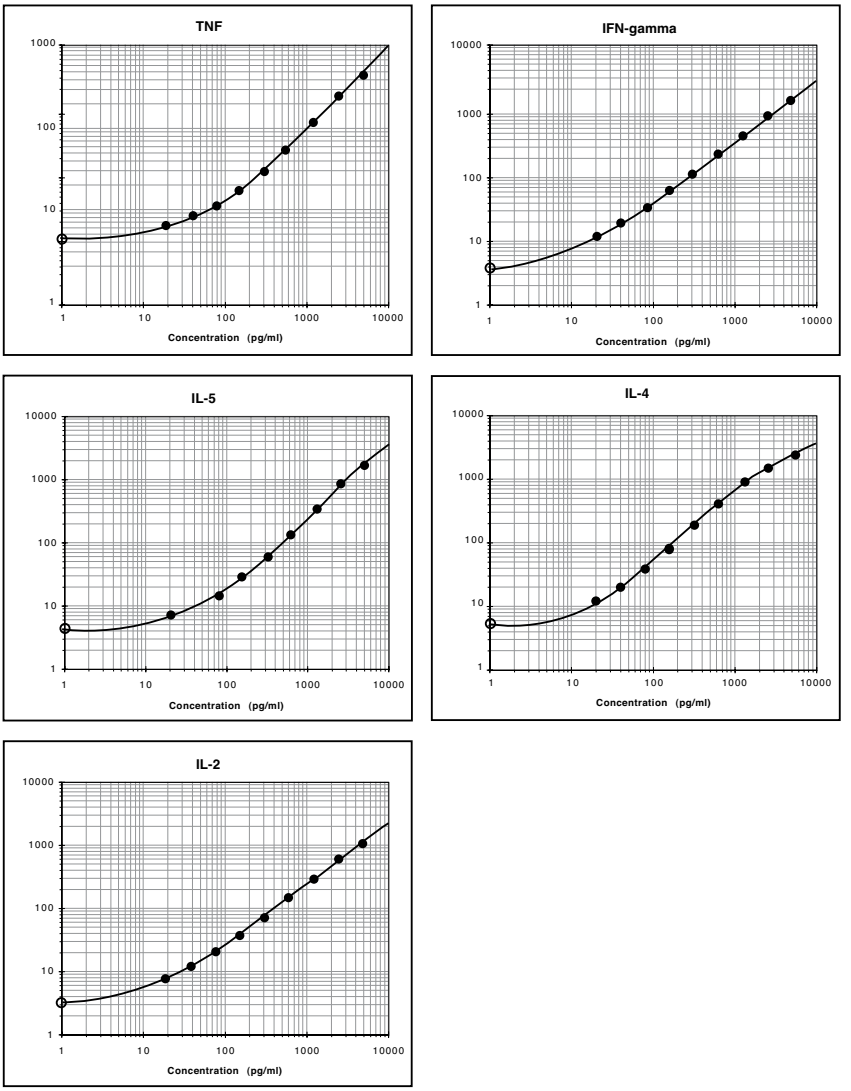


Figure 6. Example of Standard Curves

Performance

The BD CBA Mouse Th1/Th2 Cytokine assay kit has been rigorously tested for the following: theoretical limit of detection, spike recovery, spike linearity, specificity and intra- and inter-assay precision.

Theoretical Limit of Detection

The individual standard curve range for a given cytokine defines the minimum and maximum quantifiable levels using the BD CBA Mouse Th1/Th2 Cytokine assay kit (ie, 20 pg/ml and 5000 pg/ml.) By applying the 4-parameter curve fit option it is possible to extrapolate values for sample intensities not falling within the limits of the standard curve. It is up to the researcher to decide the best method for calculating values for unknown samples using this assay. The theoretical limit of detection for each cytokine using the BD CBA Mouse Th1/Th2 Cytokine assay kit is defined as the corresponding concentration at two standard deviations above the median fluorescence of 20 replicates of the negative control (0 pg/ml).

Cytokine	Median Fluorescence	Standard Deviation	Limit of Detection (pg/ml)
IL-2	5.0	0.6	5.0
IL-4	4.8	0.6	5.0
IL-5	4.4	0.7	5.0
IFN-γ	4.5	0.6	2.5
TNF	4.5	0.6	6.3

Recovery

Individual cytokine protein was spiked into various matrices at three different levels within the assay range. The cell culture media used in these experiments was not diluted before addition of the cytokine protein. The pooled mouse serum samples in these experiments were diluted 1:4 or 1:10 in Assay Diluent before addition of the cytokine protein. Results are compared with the same concentrations of the cytokines spiked in the Standard Diluent, as follows:

Cytokine	Matrix	Standard spike concentration (pg/ml)	Observed in given matrix (pg/ml)	% Recovery
IL-2	Pooled mouse sera 1:4 dilution	2500	1936.7	77%
		625	455.4	73%
		80	54.5	68%
IL-2	Pooled mouse sera 1:10 dilution	2500	1991.1	80%
		625	460	74%
		80	52.7	66%
IL-2	Cell culture media	2500	2303.4	92%
		625	592.9	95%
		80	67.7	85%
IL-4	Pooled mouse sera 1:4 dilution	2500	1002.5	40%
		625	142.7	23%
		80	18.6	23%
IL-4	Pooled mouse sera 1:10 dilution	2500	1724.3	69%
		625	243	39%
		80	27.7	35%
IL-4	Cell culture media	2500	2566.6	103%
		625	631.5	101%
		80	62.7	78%
IL-5	Pooled mouse sera 1:4 dilution	2500	2328.1	93%
		625	532.6	85%
		80	58	72%
IL-5	Pooled mouse sera 1:10 dilution	2500	2235.7	89%
		625	505	81%
		80	51.7	65%
IL-5	Cell culture media	2500	2414.8	97%
		625	592.4	95%
		80	61.8	77%
IFN-γ	Pooled mouse sera 1:4 dilution	2500	2127.7	85%
		625	475.1	76%
		80	74.1	93%
IFN-γ	Pooled mouse sera 1:10 dilution	2500	2164.6	87%
		625	546	87%
		80	71.8	90%
IFN-γ	Cell culture media	2500	2328.3	93%
		625	612.7	98%
		80	74.4	93%
TNF	Pooled mouse sera 1:4 dilution	2500	960.3	38%
		625	253.6	41%
		80	30.7	38%
TNF	Pooled mouse sera 1:10 dilution	2500	1082.7	43%
		625	273	44%
		80	34.3	43%
TNF	Cell culture media	2500	2244.1	90%
		625	572.1	92%
		80	64.6	81%

Linearity

In two experiments, the following matrices were spiked with IL-2, IL-4, IL-5, IFN-γ, and TNF and then were serially diluted with Assay Diluent.

Matrix	Dilution	Observed IL-2 (pg/ml)	Observed IL-4 (pg/ml)	Observed IL-5 (pg/ml)
Pooled mouse sera 1:4 starting dilution	1:4	3422	2421	3946
	1:8	2006	1799	2497
	1:16	1021	1052	1167
	1:32	498	506	507
	1:64	222	232	197
	1:128	105	96	85
	1:256	52	42	34
	1:512	25	17	14
	1:1024	8.1	6	2
	Slope	1.07	1.10	1.31
Cell Culture Media	Neat	4202	2797	4186
	1:2	2696	2551	2765
	1:4	1328	1705	1354
	1:8	632	814	610
	1:16	298	350	258
	1:32	135	147	117
	1:64	63	56	46
	1:128	26	23	18
	1:256	13	11	8
	Slope	1.07	1.08	1.17

Matrix	Dilution	Observed IFN-γ (pg/ml)	Observed TNF (pg/ml)
Pooled mouse sera 1:4 starting dilution	1:4	4108	2019
	1:8	2316	1131
	1:16	1159	600
	1:32	588	341
	1:64	289	192
	1:128	131	95
	1:256	66	50
	1:512	30	26
	1:1024	11	6
	Slope	1.05	0.97
Cell Culture Media	Neat	4679	4862
	1:2	2598	2741
	1:4	1326	1311
	1:8	634	576
	1:16	311	295
	1:32	148	146
	1:64	74	64
	1:128	33	21
	1:256	16	10
	Slope	1.03	1.12

Specificity

The antibodies used in the BD CBA Mouse Th1/Th2 Cytokine assay kit have been screened for specific reactivity with their specific cytokines. Analysis of samples containing only a single recombinant cytokine protein found no cross-reactivity or background detection of cytokine in other Capture Bead populations using this assay.

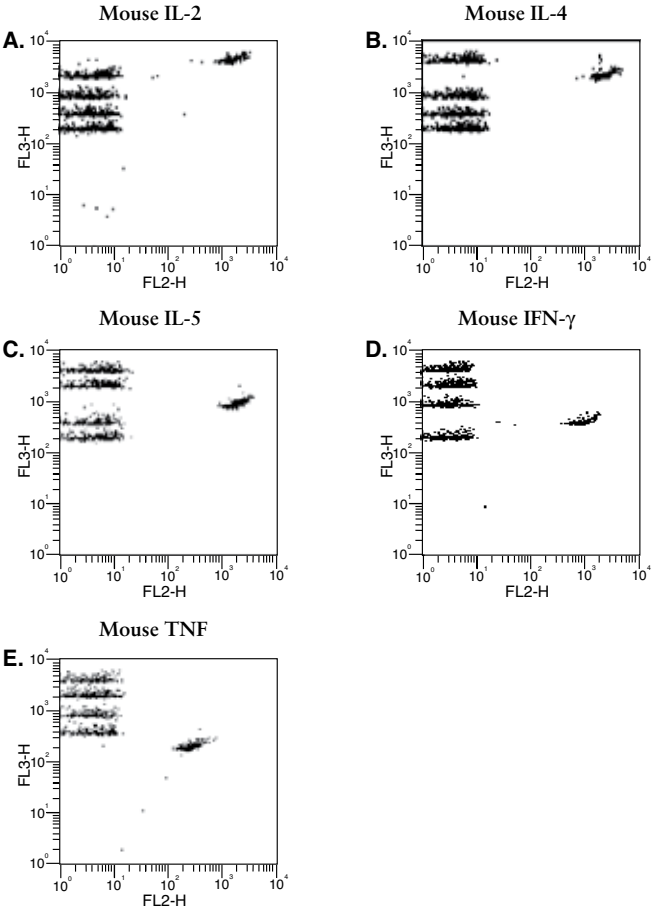


Figure 7. BD CellQuest Data for Detection of Individual Cytokines

Precision

Intra-assay: Ten replicates of each of three different levels of IL-2, IL-4, IL-5, IFN-γ, and TNF were tested.

Cytokine	IL-2			IL-4		
Actual Mean Conc. (pg/ml):	58	469	2219	59	557	2648
SD	3	26	62	2	37	73
% CV	6%	6%	3%	4%	7%	3%

Cytokine	IL-5			IFN-γ		
Actual Mean Conc. (pg/ml):	50	463	2493	73	594	2565
SD	4	26	83	2	16	108
% CV	9%	6%	3%	3%	3%	4%

Cytokine	TNF		
Actual Mean Conc. (pg/ml):	59	482	2138
SD	7	30	57
% CV	11%	6%	3%

Inter-assay: Three different levels of IL-2, IL-4, IL-5, IFN-γ, and TNF (80, 625 and 2500 pg/ml) were tested in four experiments conducted by four different operators.

Cytokine	IL-2			IL-4		
Number of Replicates:	8	8	8	8	8	8
Actual Mean Conc. (pg/ml):	63	544	2362	61	565	2634
SD	4.9	44	218	4	58	283
% CV	8%	8%	9%	7%	10%	11%

Cytokine	IL-5			IFN-γ		
Number of Replicates:	8	8	8	8	8	8
Actual Mean Conc. (pg/ml):	54	525	2376	77	602	2419
SD	7	56	294	6	44	194
% CV	14%	11%	12%	8%	7%	8%

Cytokine	TNF		
Number of Replicates:	8	8	8
Actual Mean Conc. (pg/ml):	73	551	2341
SD	13	43	198
% CV	18%	8%	8%

Note: The number of replicates refers to the total number of assay tubes tested at a given concentration of protein.

Troubleshooting Tips

Problem	Suggested Solution
Variation between duplicate samples.	Vortex Capture Beads before pipetting. Beads can aggregate.
Low bead number in samples.	Avoid aspiration of beads during wash step. Do not wash or resuspend beads in volumes higher than recommended volumes.
High background.	Test various sample dilutions, the sample may be too concentrated. Remove excess Mouse Th1/Th2 PE Detection Reagent by increasing the number of wash steps as the background may be due to non-specific binding.
Little or no detection of protein in sample.	Sample may be too dilute. Try various sample dilutions.
Less than five bead populations are observed during analysis or distribution is unequal.	Ensure that equal volumes of beads were added to each assay tube. Vortex Capture Bead vials before taking aliquots. Once Capture Beads are mixed, vortex to ensure that the beads are distributed evenly throughout the solution.
Debris (FSC/SSC) during sample acquisition.	Increase FSC threshold or further dilute samples. Increase number of wash steps if necessary.
Overlap of bead population fluorescence (FL3) during acquisition.	This may occur in samples with very high cytokine concentration. Ensure that instrument settings have been optimized using the Cytometer Setup Beads.
Standards assay tubes show low fluorescence or poor standard curve.	Check that all components are properly prepared and stored. Use a new vial of standard with each experiment and once reconstituted, do not use after 12 hours. Ensure that incubation times were of proper length.
All samples are positive or above the high standard mean fluorescence value.	Dilute the samples further. The samples may be too concentrated.

Note: For best performance, Vortex samples immediately before analyzing on a flow cytometer.

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Notes

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