BD FACSDiscover™ S8 Cell Sorter Quick Reference Guide

This guide contains instructions for using the BD FACSDiscover™ S8 Cell Sorter with BD CellView™ Image Technology and BD SpectralFX™ Technology. See the user's guide for additional information.

Workflow overview

The following figure shows a typical daily workflow when using the BD FACSDiscover™ S8 Cell Sorter.

Before you begin

- Verify the sheath tank is full, and the waste tank is empty.
- Prepare the BD FACSDiscover™ Setup Beads, BD FACS™ Accudrop Beads and BD CellView™ Calibration Beads according to the package insert.
- Prepare the single-stain controls for your experiment.

Start up system

1. Turn on the source of air pressure and verify that the output is 80–95 psi.
2. If you work with a biological safety cabinet (BSC) or an aerosol management system (AMO), ensure it is turned on for 3 minutes before powering on the cytometer.
3. Power on the cytometer.
4. Power on the workstation and log into Microsoft® Windows®.
5. Open BD FACSChorus™ Software.

Startup system, continued

1 Fluidics Startup 2 Cleaning 3 Sort Nozzle 4 Setup and QC 5 Image Calibration 6 Drop Delay

6. Select a startup process and follow the prompts on the screen.

Cytometer Connection: Connected
Sheath Tank: 19 hr 39 min remaining
Waste Tank: OK

Last Shutdown: 03/15/2023 03:26 PM Type: Daily
Last Fluidics Startup: 03/16/2023 08:58 AM Type: Daily

Run Daily Fluidics Startup | Run Extended Fluidics Startup | Skip

Click **Run Daily Fluidics Startup** after a daily shutdown.

Click **Run Extended Fluidics Startup** after a long-term shutdown.

7. Select **Flow Cell Clean** and follow the prompts on the screen.

Select the cleaning that you want to run.

Prepare for Aseptic Sort
Cleans the sheath and sample paths with bleach, DI water, and ethanol.
Last Run: 02/20/2023 03:52 PM

Flow Cell Clean
Cleans the sample path and fills the flow cell with DI water. Run this procedure when poor optical performance indicates that additional cleaning is needed.
Last Run: 03/16/2023 09:00 AM

Skip

8. Remove the closed-loop nozzle and insert a sort nozzle.

9. Perform a Setup and QC.

Verify the bead lot.

Select the type of QC.

Load a tube of BD FACSDiscover™ Setup Beads and click **Run.**

Startup system, continued

10. If needed, perform an Image Calibration.

Click **System** to view the Image Calibration status.

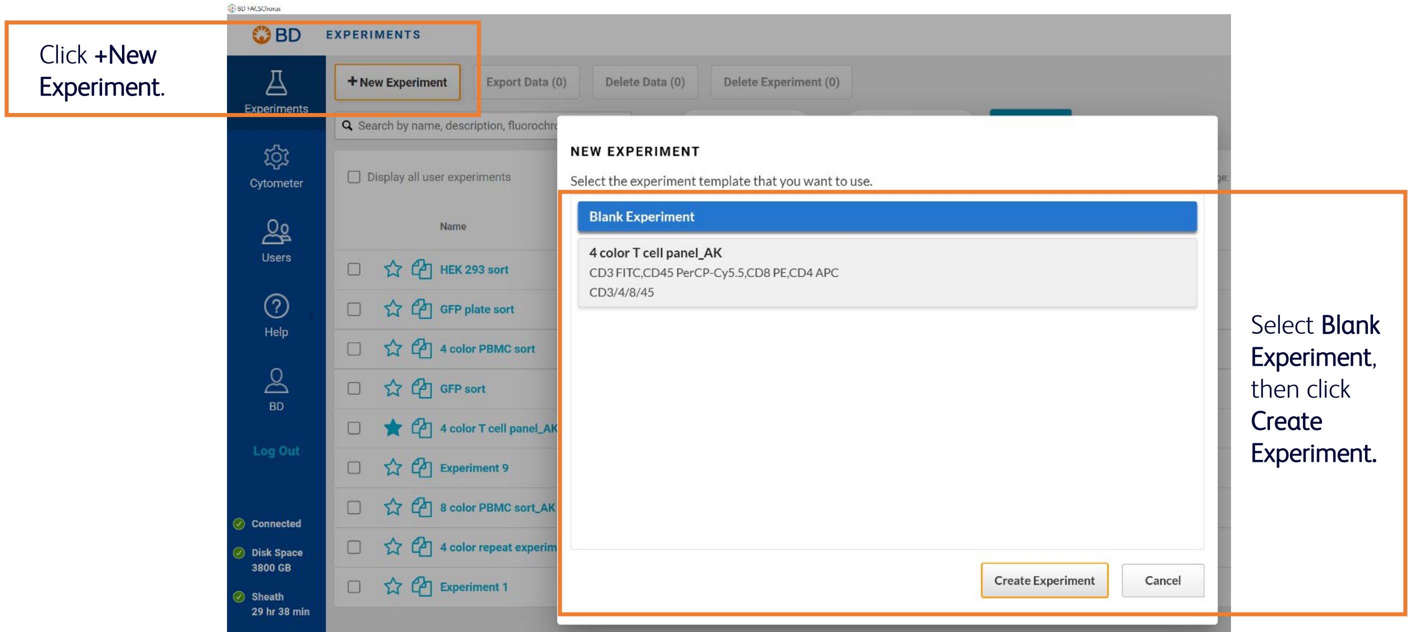
Run a tube of BD CellView™ Calibration Beads if the Image Calibration is outdated. Click **Skip** if the Image Calibration status is green.

11. Perform a Drop Delay setup.

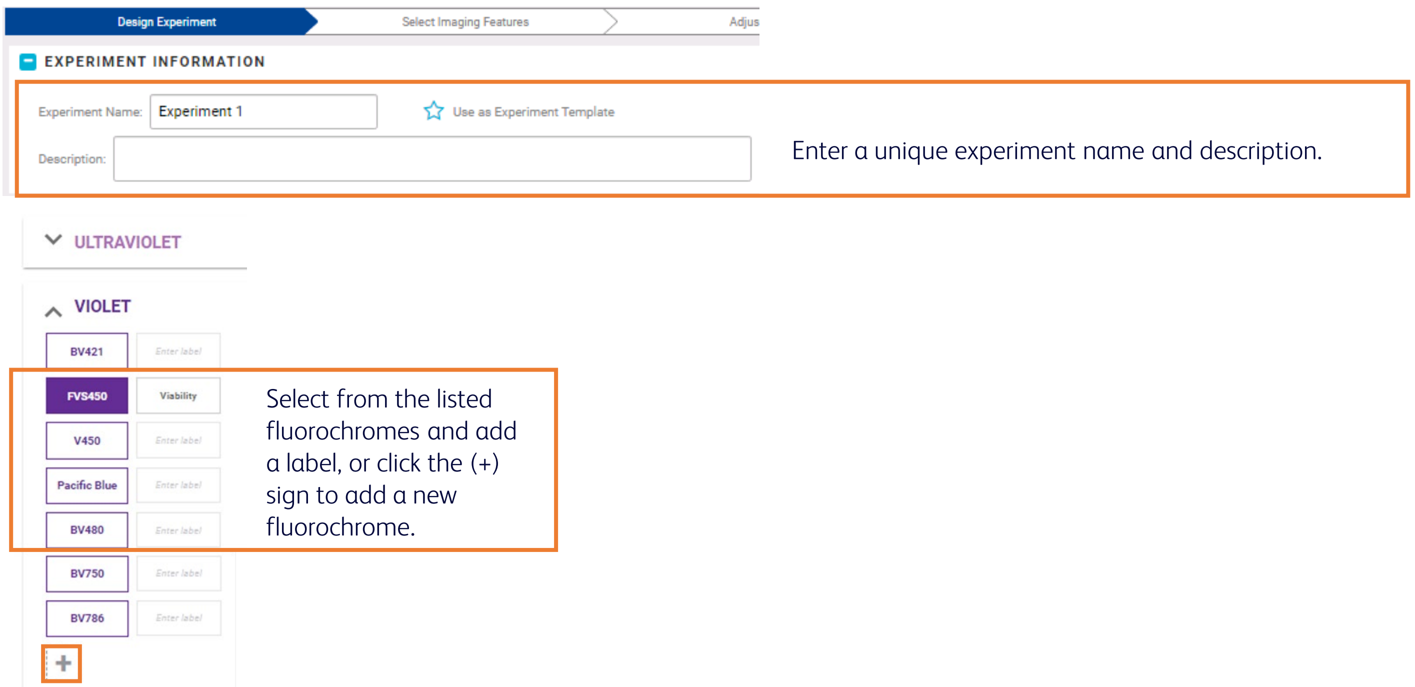
Run a tube of BD FACS™ Accudrop Beads.

Set up experiment

1. Create a new experiment on the Experiments page.

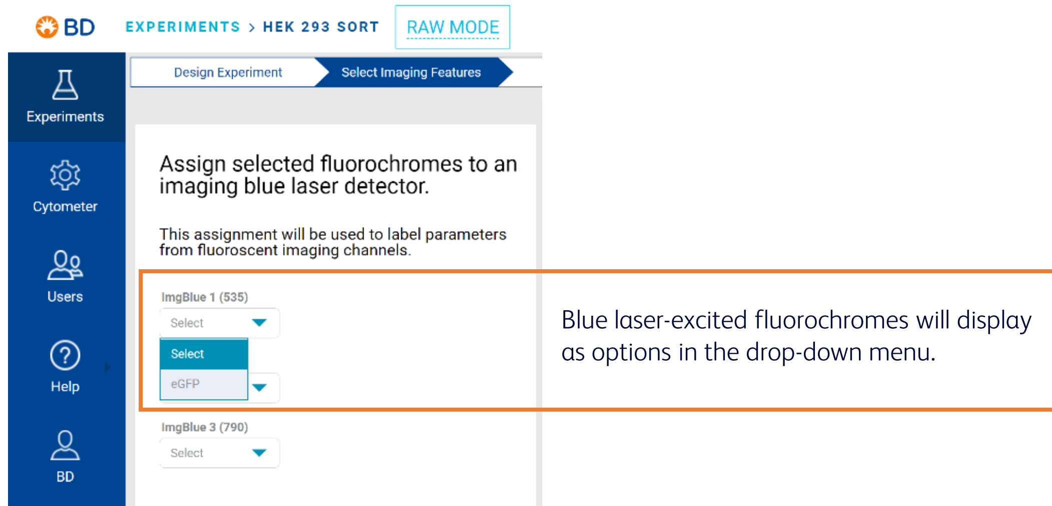


2. Enter the experiment information, dyes and labels on the Design Experiment page.



Set up experiment, continued

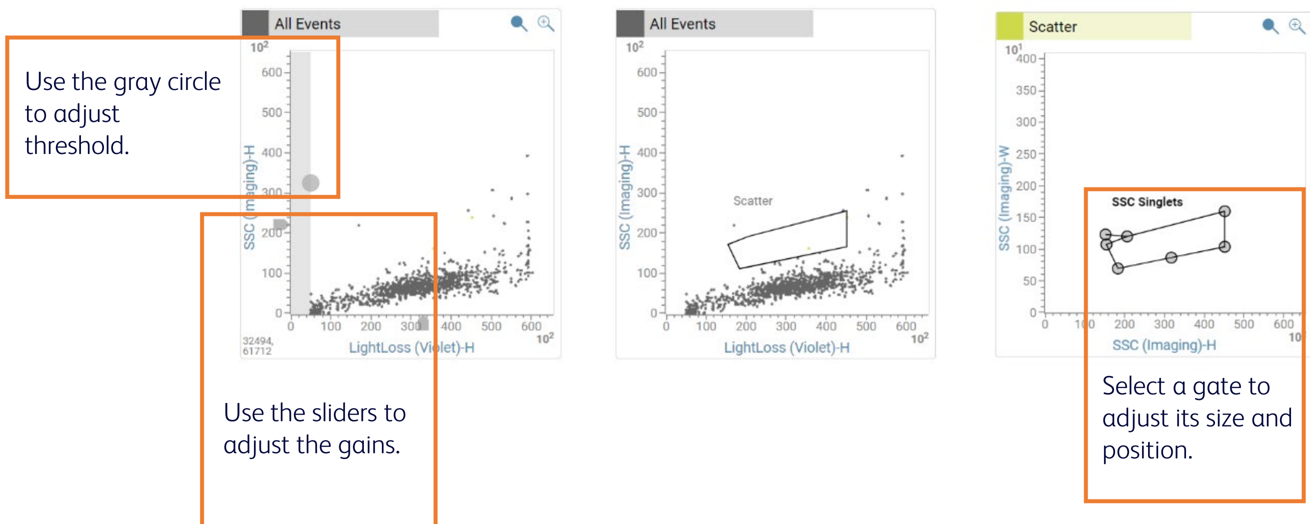
3. Enable fluorescence imaging by assigning dyes to the imaging detectors.



Establish settings

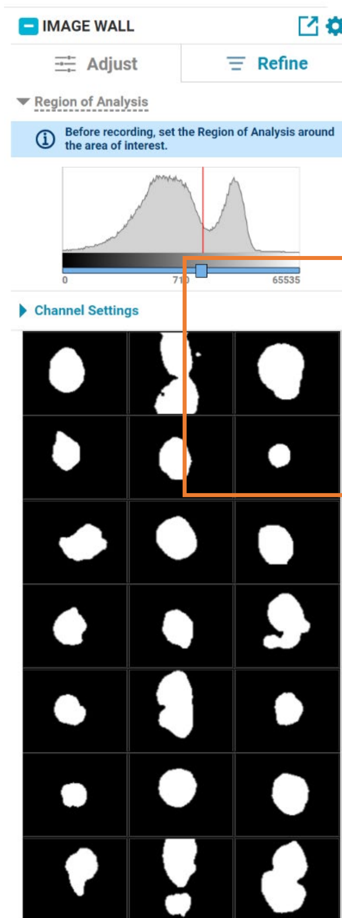
1. On the Adjust Gains page, load a tube of brightly stained cells.

2. Adjust the scatter gains, threshold, and gates to encompass cells of interest.



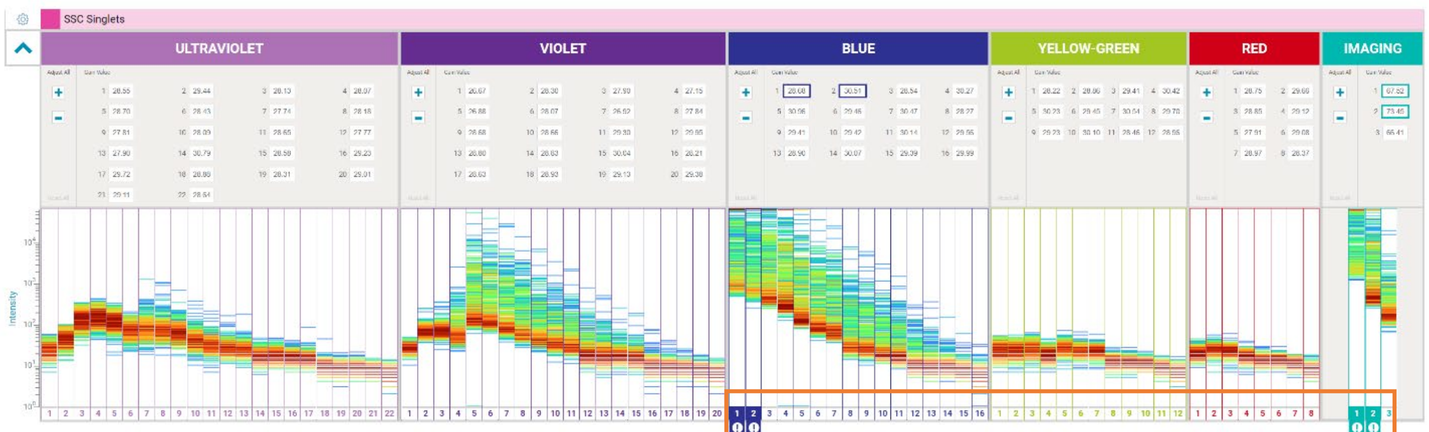
Establish settings, continued

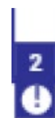
3. On the Image Wall, adjust the Region of Analysis (ROA).



Adjust the ROA slider until the white area in the images completely encompasses the particles of interest while minimizing background noise.

4. Use the spectral plot to adjust the gains, if needed.





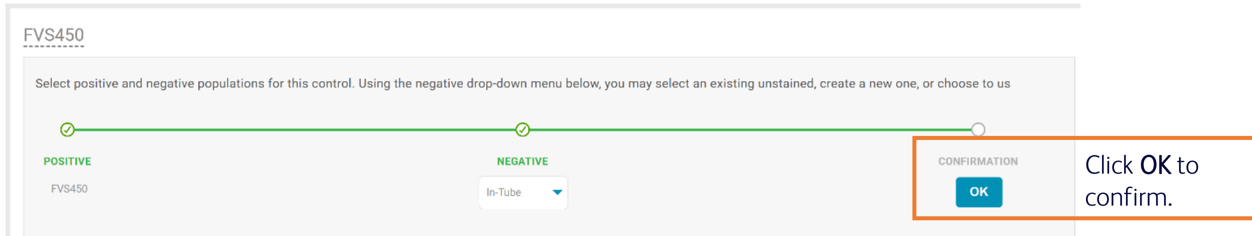
Saturated detectors will be identified with the (!) icon. Lower the gains for these detectors until the icon disappears.

Establish settings, continued

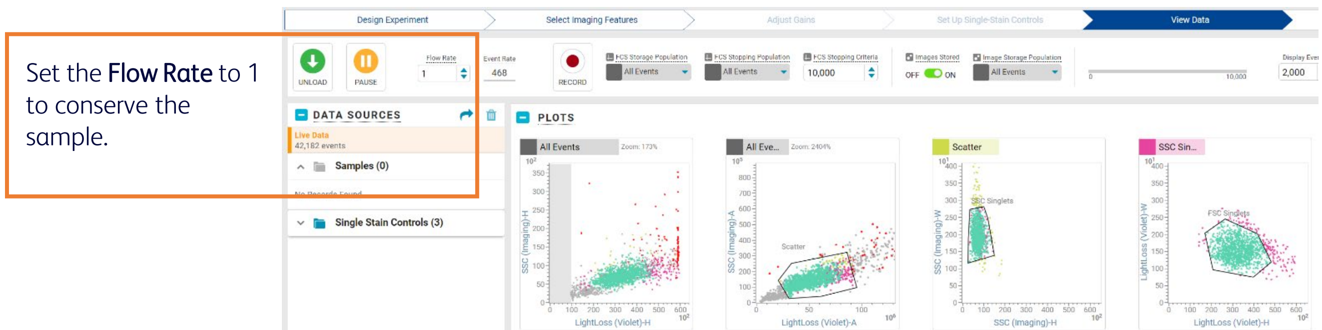
5. On the Set Up Single-Stain Controls page, record data for each control tube.

   NOTE The Region of Analysis must be appropriately set for the particle type before recording.

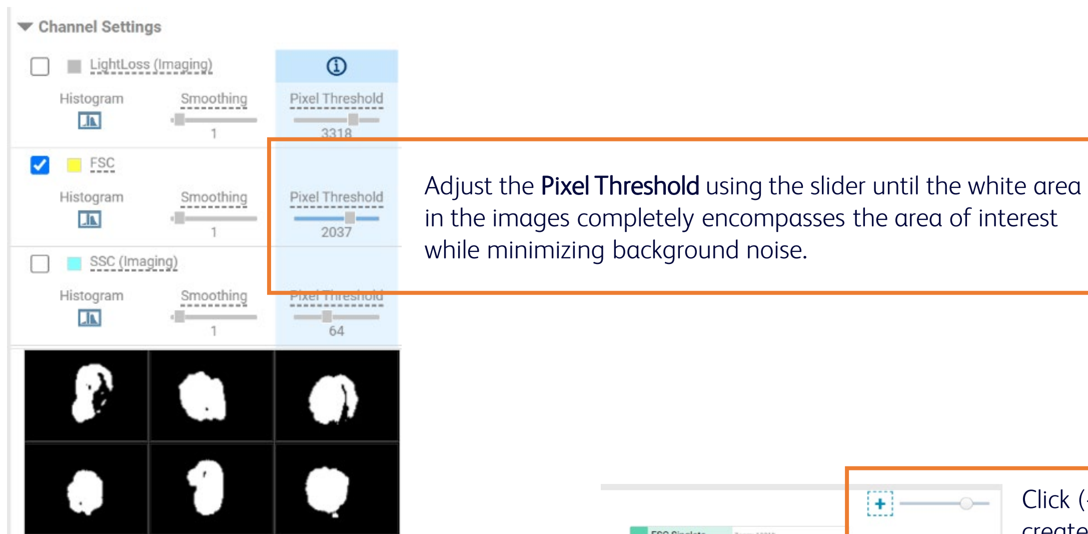
6. Adjust the gates for each parameter.

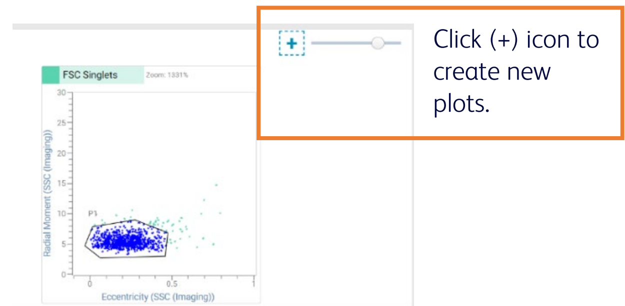


7. On the View Data page, load a sample tube and adjust scatter and singlet gates to encompass cells of interest.



8. Use the image wall to adjust the Region of Analysis and Pixel Threshold for each imaging detector.





9. Record a pre-sort data file and then name the file.

10. Create new plots and gates to identify populations of interest.

Establish settings, continued

11. Use the image wall to adjust the channel settings for each imaging detector.

Channel Settings

LightLoss (Imaging)

Color | Histogram

Select the color box.

Use the smoothing slider to reduce image blur.

Smoothing 1

Pixel Threshold 3003

Minimum 46464 | Gamma 1.00 | Maximum 55202

Adjust the minimum (red) and the Maximum (blue) bars around the signal peak.

Adjust the Gamma (black circle) as needed.

Sort and analyze

1. In the Set Up Sort page, determine the collection setup and the populations in the sample to be sorted.

COLLECTION SETUP

Format: 4-Way Tube

Volume: 5.0 mL

Sort Mode: Purity

Select the format and the volume of the collection device.

Select the **Sort Mode**: Yield, Purity, or Single Cell.

a. If you are performing a tube sort:

SORT SETUP

Tube	1	2	3	4
Initial Buffer Volume:	0.50 mL	0.50 mL	0.50 mL	0.00 mL
Number of Events:	50,000	50,000	50,000	781,250
	Max: 703,125 events	Max: 703,125 events	Max: 703,125 events	Max: 781,250 events

Specify the buffer volume and set the target number of events to be sorted for each tube.

Assign a sort population by clicking a tube and selecting the population that you want.

Punctate | Diffuse | Intermed

Population Hierarchy: All Events, Saturated, Unsaturated, Scatter, SSC Singlets, FSC Singlets, E/RM Singlets, Viable bright ..., Punctate, Intermed, Diffuse

Assign populations to tubes by selecting the tube, then the population in the hierarchy.

Sort and analyze, continued

b. If you are performing a plate sort:

Select the **BD-Defined Plate** type, **Plate Name**, and **Sort Mode**.

Assign the sort wells by clicking each well, dragging across a group of wells, clicking the letter or number for a row or column or Select All. You can also select non-contiguous wells by using Ctrl+click.

Specify the buffer volume and set the target number of events to be sorted for each well.

Select the sort population from the Population Hierarchy.

2. Install the collection device into the sort collection chamber and close the door.

3. On the Sort page, load the sort sample and start the sort.

Sort and analyze, continued

4. Monitor the sort as it progresses.

Monitor the event rate.

Monitor the sort efficiency and sort rate.

Verify that the sort gates are still capturing the appropriate populations throughout the sort.

5. When the sort ends or is stopped, name the sort report.

6. On the View Data page, analyze the sort purity.

TIP Perform a backflush between tubes to reduce carryover.

BACKFLUSH Light Agitation OFF Temperature OFF

Manage data

1. On the View Reports page, view and export reports.

Design Experiment | Select Imaging Features | Adjust Gains | Set Up Single-Stain Controls | View Data | Set Up Sort | Sort | View Reports

Select Sort Report: Tube sort | Export Report

Tube sort

CYTOMETER INFO

CYTOMETER SETTINGS

Gains

Threshold: LightLoss (Violet) @ 9788

Detector	Gains
SSC (Imaging)	40.17
FSC	18.44
LightLoss (Imagin...	10.12
LightLoss (Violet)	12.86

Spectral Unmixing: Spillover Values

Export as CSV

Fluorochromes x Detectors	UV1	UV2	UV3	UV4	UV5	UV6	UV7
FVS450	0.413	1.242	4.560	11.143	11.956	6.710	4.972
eGFP	0.044	0.081	0.253	0.300	0.230	0.229	1.269
Autofluoresce...	14.877	24.694	75.605	100.000	92.407	60.288	67.955

Manage data, continued

2. On the Experiments page, export FCS files, images, and index sort data.

3. Delete unneeded experiments.

Export Data, Delete Data, or Delete Experiment.

Select an experiment.

Shut down system

1. Clean the sample line.

2. On the Cytometer page, perform either a daily or long-term shutdown.

Perform a Long-Term Shutdown if the instrument is idle for over two days. Otherwise, perform a Daily Shutdown.

3. Power off the cytometer and workstation.

This material is for training purposes.
For Research Use Only. Not for use in diagnostic or therapeutic procedures.

BD Life Sciences, San Jose, California, 95131, USA

bdbiosciences.com

BD, the BD Logo, BD FACS, BD CellView, BD FACSChorus, BD FACSDiscover, BD FACSFlow and BD SpectralFX are trademarks of Becton, Dickinson and Company or its affiliates. ©2023 BD. All rights reserved. NPM-1354

BD