

BD Accuri™ C6 Flow Cytometer

Mirko Corselli, PhD BD Biosciences Senior Scientist

The BD Accuri C6 Flow Cytometer System BD

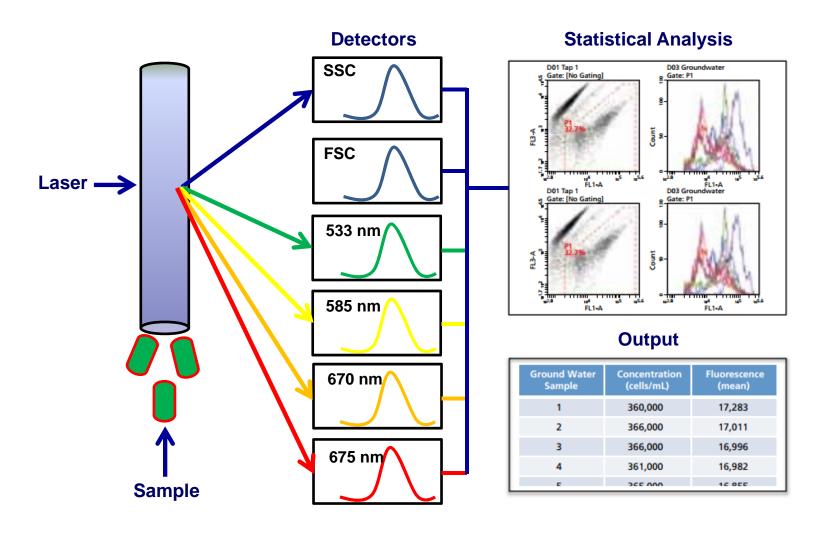


- An affordable, full-featured, easy-to-use flow cytometer
- Two lasers and six detectors



BD Accuri C6 Flow Cytometer





Flow Cytometry within Reach™

Advantages of Pre-optimized Detector Settings



- Greatly reduces the risk of lost data due to improper setup
- Saves time and sample
- No specialist training or dedicated operator required
- Predictable, reproducible analysis relative to the sample type and application

Enhanced Sample Handling



- Direct volume measurement
- Many types of sample tubes may be used.
 - Flow cytometry tubes
 - Microcentrifuge tubes
 - Ninety-six-well plates with the BD CSampler™ accessory
- Open system conducive to kinetic studies
- BD CSampler™ accessory for automated sample introduction







Intuitive Software



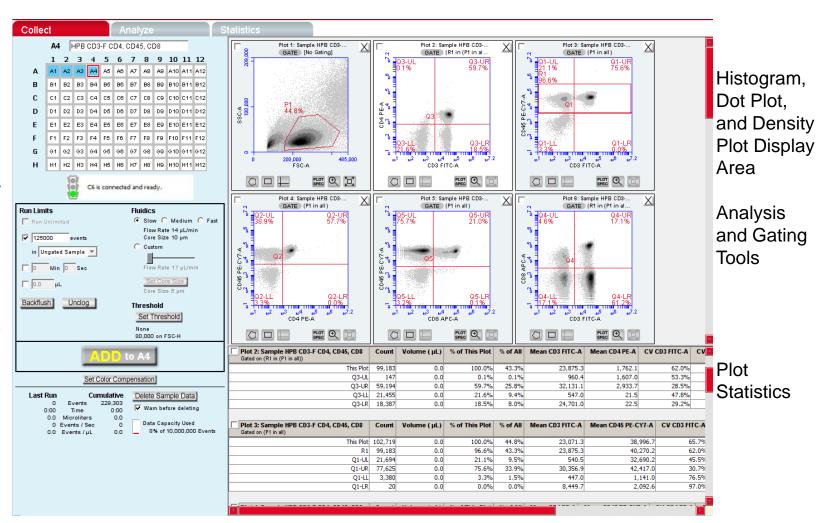
Sample Grid

Cytometer Status

> Fluidics Controls

Run Criteria

Real-Time Updates

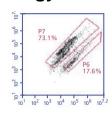


A Versatile Instrument for Broad Applications



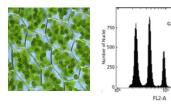
Microbiology





- Aquatic microbiome analysis
- Biofuel research
- Bacteria viability and concentration

Plant Biology

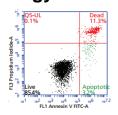


DNA content



Cell Biology

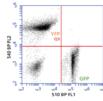




- Apoptosis
- Proliferation
- Immunophenotyping

Fluorescent Protein Analysis





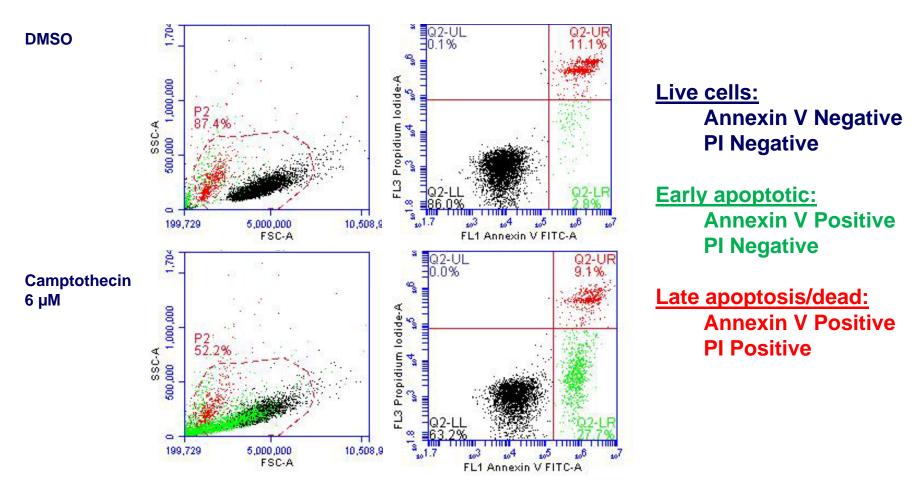
- GFP,YFP
- mCherry®, RFP
- mOrange®, dTomato®

Flow Cytometry within Reach™

The BD Accuri™ C6 Personal Flow Cytometry Tour

Apoptosis: Annexin V

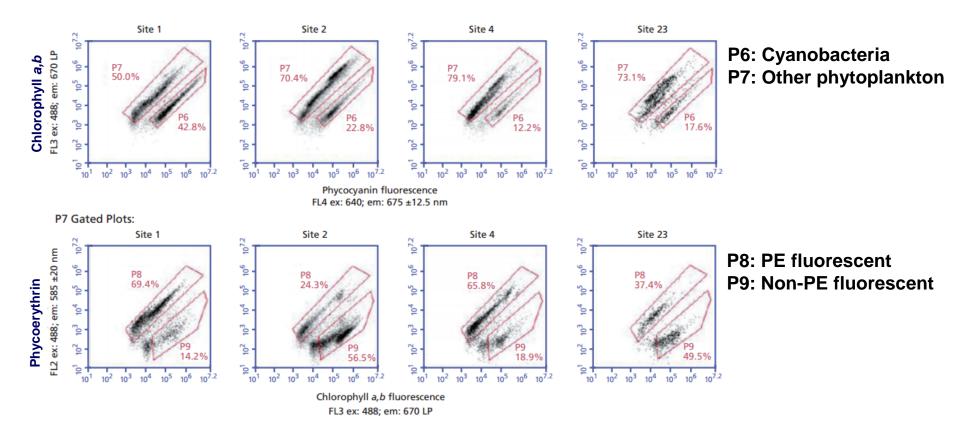




Jurkat cells were treated with DMSO or camptothecin for 4 hours.

Analysis of Aquatic Samples from Saginaw Bay: Fluorescence





Four types of phytoplankton were identified by fluorescence characteristics.

Data courtesy of J.D. Bressie, PhD, NOAA, Seattle, WA

Kits and Templates on the BD Accuri C6



Category	Product Information Sheet	Brand	Kit	Cat. No.	Template
Cell Biology	BD Apoptosis Kits and Templates	BD Pharmingen™	Annexin V FITC Apoptosis Detection Kit II	556570	Download
		BD Pharmingen™	Annexin V PE Apoptosis Detection Kit I	559763	Download
		BD™	MitoScreen (JC-1) Kit	551302	Download
		BD Pharmingen™	Caspase-3 PE Assay Kit	550914	Download
		BD Pharmingen™	Caspase-3 FITC Assay Kit	550480	Download
	BD Cell Cycle and DNA Kits and Templates	BD Cycletest™ Plus	DNA Reagent Kit	340242	Download
		BD Pharmingen™	FITC BrdU Flow Kit	559619	Download
		BD Pharmingen™	APC BrdU Flow Kit	552598	Download

BD Accuri C6 Promotion and Personal Flow Cytometry Tour





Note:

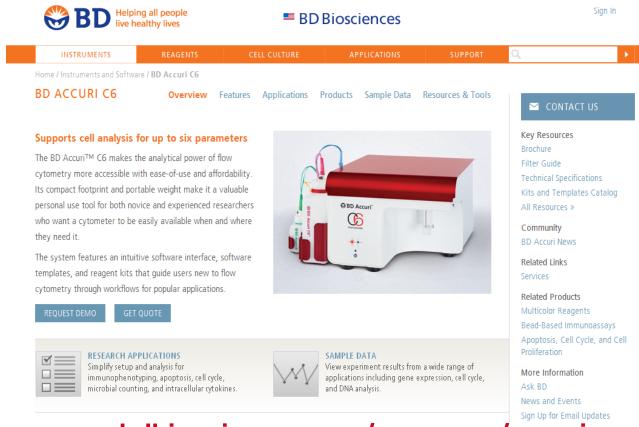
US Region Only

Promotion Period: Oct 1, 2014 - Dec 31, 2014

The BD Accuri C6 Personal Flow Cytometry Tour

- Introduction to Flow Cytometry
- Cancer and Cell Biology Applications
- Microbial Analysis

For Additional Information...



www.bdbiosciences.com/resources/accuri

Technical Support:

Ph: 877-232-8995, Prompt 3, 2

email: ResearchApplications@bd.com

Class 1 Laser Product.

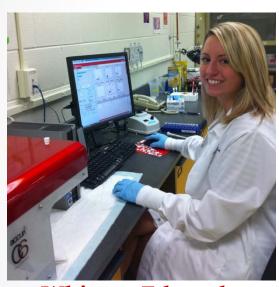
For Research Use Only. Not for use in diagnostic or therapeutic procedures.

Living Colors® (including mCherry, mOrange, and dTomato dyes) is a registered trademark of Clontech.

BD, BD Logo and all other trademarks are property of Becton, Dickinson and Company. © 2014 BD 23-16980-00



Flow cytometry in undergraduate education



Whitney Edwards



Samantha Scott, Talbot Weston, Sarah Murphy, Melanie Gubbels Bupp – Autumn Immunology Conference 2014

Melanie Gubbels Bupp, Assistant
Professor of Biology



General Advice on using cytometry in higher education

- Use the technology for projects that allow students to...
 - o more firmly grasp basic concepts
 - o test something of interest to them
- Only tell them what they need to know about the technology WHEN they need to know it.
 - o Front-loading a lot of technical information is overwhelming and often, ineffective

Examples of using cytometry at R-MC

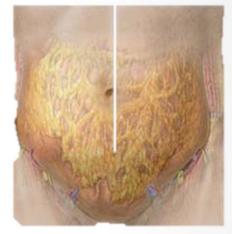
- Peritonitis Lab in Immunology course for biology majors [simplified version taught in a freshman mixed (majors/non-majors) lab]
 - Phagocytosis Lab in Immunology course for biology majors
 - Independent research projects for undergraduate students



Introduce students to the lab

Thioglycollate-Induced Peritonitis: Recruitment of leukocytes from the circulation, and their subsequent influx into the sites of inflammation is critical for host defense and wound healing. This is a multistep process, which is regulated, in part, by adhesion molecules and chemokines that are upregulated during inflammation. An intra-peritoneal injection of thioglycollate generates local inflammation and initiates the migration of inflammatory cells to the site of inflammation. Thus, thioglycollate-induced peritonitis in mice mimics an acute inflammatory response in the peritoneum.

Peritoneum



Learning Objectives

- Be able to identify particular white blood cells (lymphocytes, monocytes, neutrophils) in a blood smear.
- Explain how and why blood differentials are taken
- Describe the chronological order in which immune cells arrive at sites of inflammation, such as the peritoneal cavity in thioglycollate-treated mice.



Peritonitis Lab Timeline



Inject mice with TG (1 Finish blood hr timepoint); Blood Group differential analysis; differentials on all TG Learn how to perform presentations on acquire cells on BD samples; count & stain and read blood lab + discussion Accuri C6; data peritoneal cells from differentials questions analysis control and TG mice (untreated mice) **Weekly Lab Session**

Inject mice with 6% thioglycollate (TG) in PBS or control PBS to induce peritoneal inflammation



R-MC Immunology students acquiring their samples

Victoria Robinson



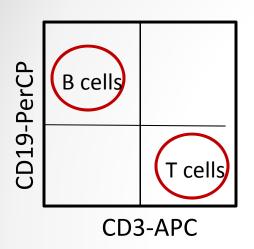
Lauren Philips, Jane Oh, & Casey Kaufman

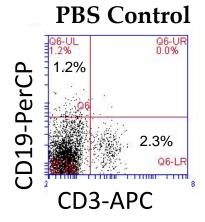


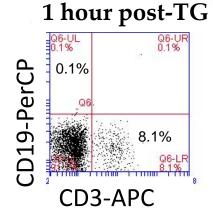


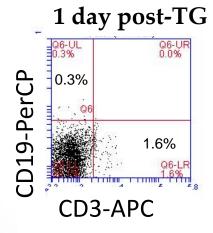
Explaining how cytometry works Labeled CD8+ DP antibody specific to CD4 Each dot is a cell Labeled antibody specific to CD8 CD4+ Thymus DN CD4 **Flow Cytometer** CD4

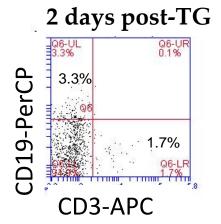
Example student data





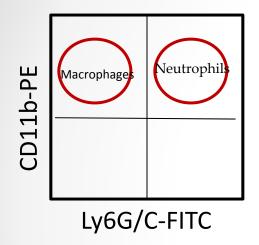


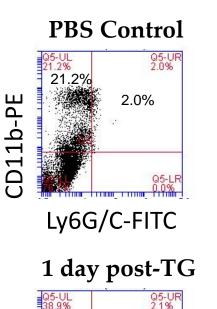


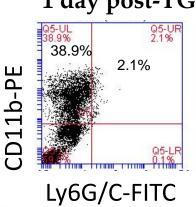


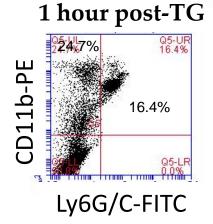


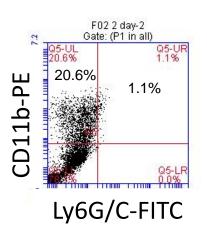
Example student data













Example discussion questions for student peritonitis presentations

- What types of junctions exist between endothelial cells lining blood vessel walls in the non-inflamed, resting condition? How do these junctions change during local inflammation?
- Relate your findings from the blood differentials with your findings from the flow cytometry data. Do the two sets of data "paint the same picture"? Why or why not?
- Map the route newly developed neutrophils must take to enter the inflamed peritoneal cavity. Begin in the bone marrow and end with the peritoneal cavity.

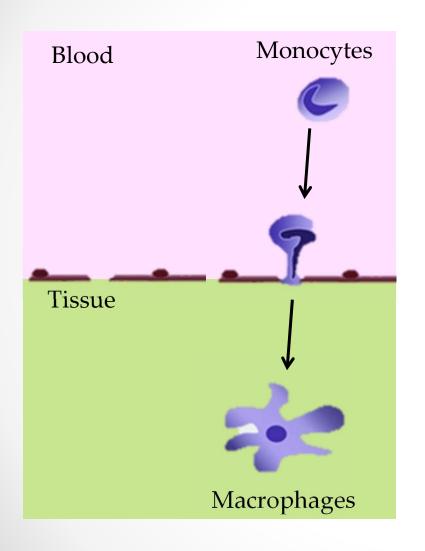


Examples of using cytometry at R-MC

- Peritonitis Lab in Immunology course for biology majors [simplified version taught in a freshman mixed (majors/non-majors) lab]
- Phagocytosis Lab in Immunology course for biology majors
 - Independent research projects for undergraduate students



Introduce students to the lab

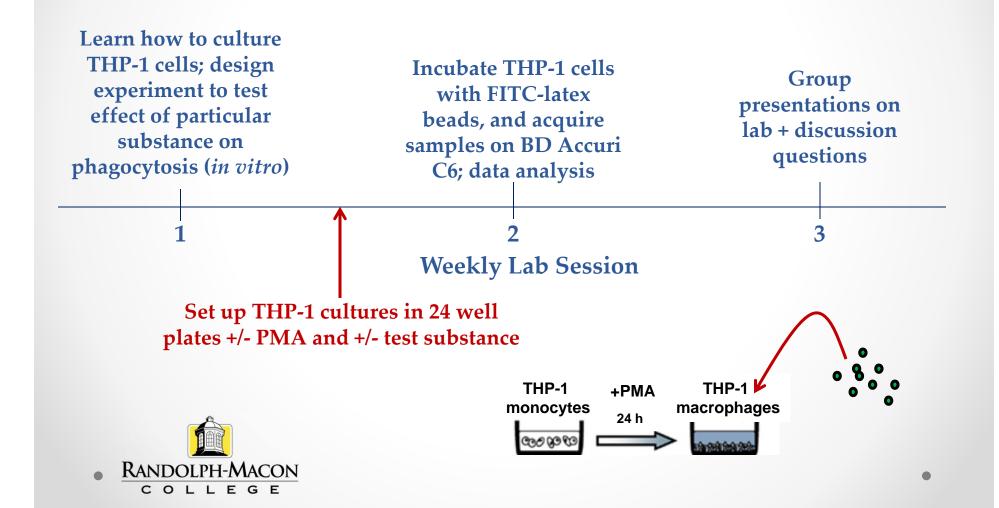


Learning Objectives

- Compare and contrast monocytes and macrophages
- Evaluate flow cytometry data regarding the ability of cells to phagocytose fluorescently tagged antigens
- Be able to design an experiment to test the impact of various compounds on phagocytosis

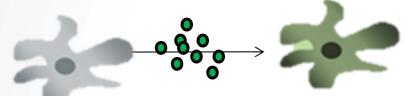


Phagocytosis Lab Timeline

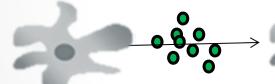


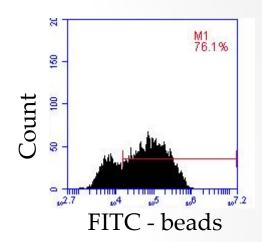
Explaining how cytometry works -- histograms

Lots of phagocytosis

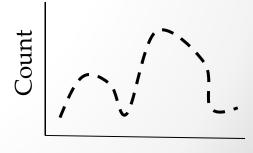


Very little phagocytosis





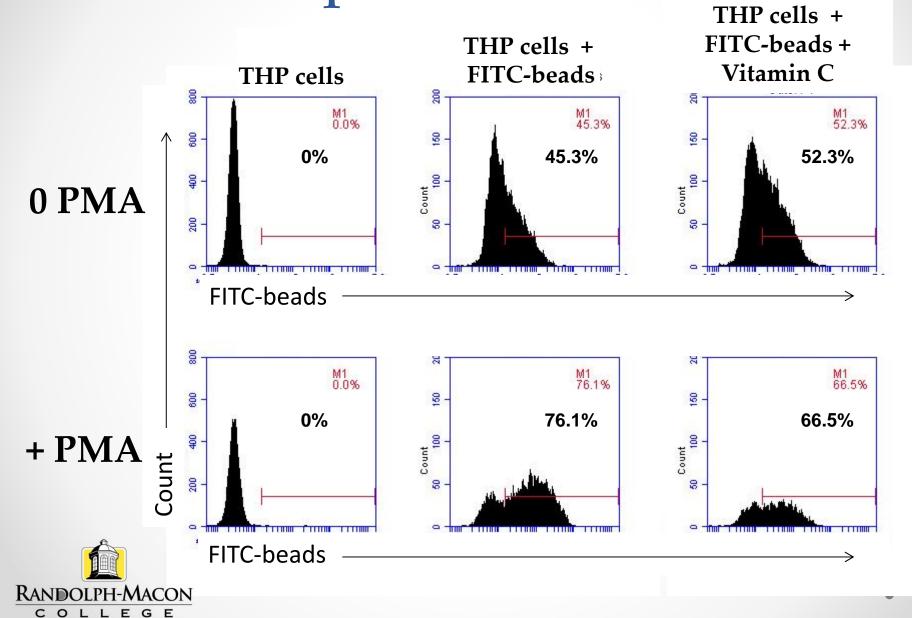
Flow Cytometer



FITC-beads



Example student data



Examples of using cytometry at R-MC

- Peritonitis Lab in Immunology course for biology majors [simplified version taught in a freshman mixed (majors/non-majors) lab]
- Phagocytosis Lab in Immunology course for biology majors
- Independent research projects for undergraduate students

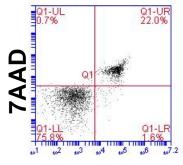


Students at R-MC also use the cytometer in independent research projects

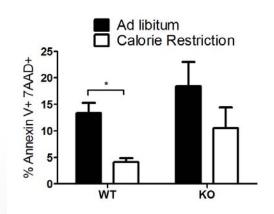


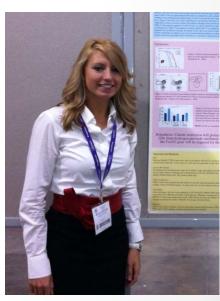
Seth Litvin

Apoptosis Markers



Annexin V - PE





Whitney Edwards



Students at R-MC also use the cytometer in independent research projects

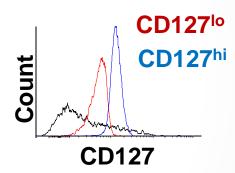
Sarah Murphy



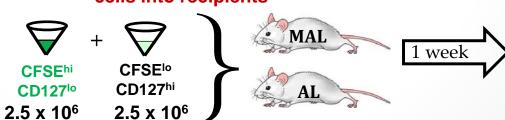


Isolate Naïve CD8+ T cells from Donor mice

Sort them on CD127 expression



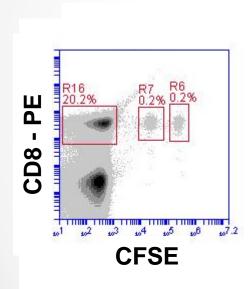
Adoptively Transfer cells into recipients

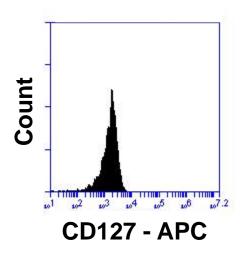


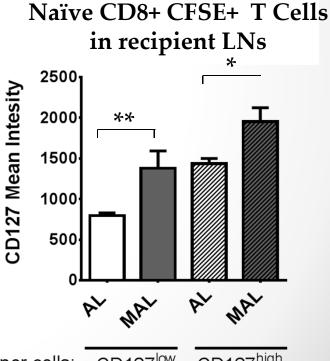
Assess total numbers and CD127 expression of CFSE+ cells by flow cytometry



Students at R-MC also use the cytometer in independent research projects









Donor cells: CD127^{low}

Thank you to R-MC students participating in labs and independent research

- Whitney Edwards
- Erica Horseman
- Rebecca Davis
- Seth Litvin
- Victoria Robinson
- Josh Anoff
- Brittany Mihalcoe
- Alex Koppleman
- Samantha Scott
- Sarah Murphy
- Talbot Weston



Talbot Weston, Sarah Murphy, and Samantha Scott



Integrating *Microbial* Flow Cytometry Into Education

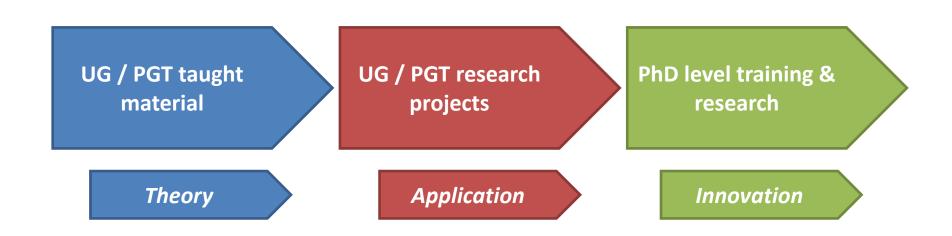
Tim W Overton
Bioengineering, School of Chemical Engineering
University of Birmingham

t.w.overton@bham.ac.uk @overtonlab

How can we get FCM applied in microbiology / microbial biotechnology?

 Collaboration between FCM specialists and microbiology / bioprocessing researchers

Training at University level

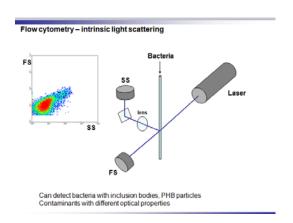


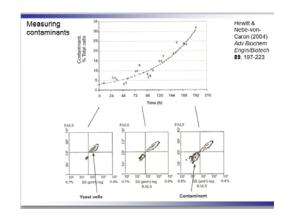
Teaching fermentation

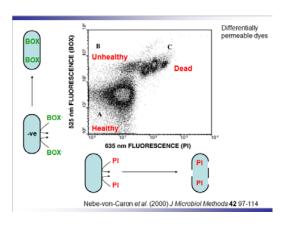
- MSc Biochemical Engineering
 - Fermentation and cell culture for production of biopharmaceuticals
 - Downstream processing of biopharmaceuticals
 - Systems and synthetic biology
 - Pharmaceutical, food and business themes
 - Research project

Teaching analysis of fermentation

- Theory sessions on analytical techniques
 - Online versus offline
 - Real time / non-real time
 - Bulk versus single cell
 - Direct observations
- Theory and advantages of FCM





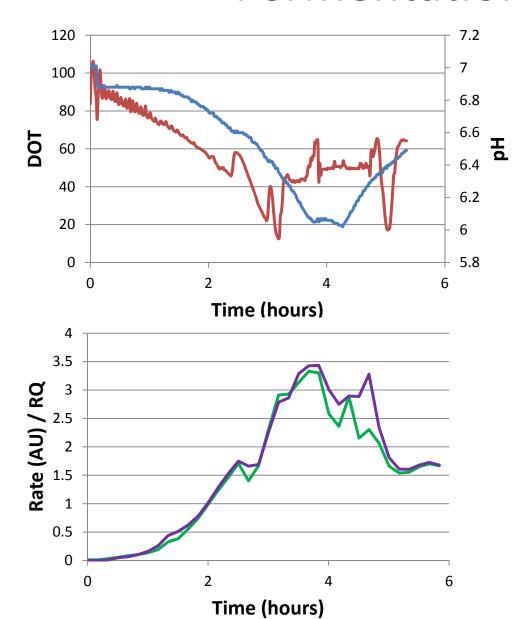


Lab-scale fermentation

- Growth of *E. coli* in 5 litre bioreactors
- Analysis:
 - Traditional techniques:pH, DOT, offgas
 - Biomass measurements:
 - Optical density
 - Colony forming units
 - Dry cell weight
 - FCM



Fermentation data

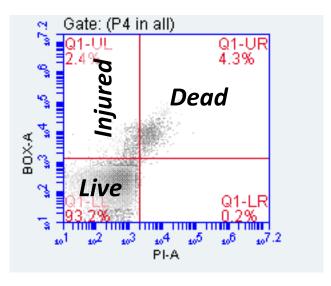


Traditional techniques:
Dissolved O₂ and pH
Basics of growth and
physiology

O₂ consumption, CO₂ evolution Basics of growth and physiology

FCM with physiology dyes

1 hour



DiBac₄(3) (Bisoxanol; BOX)

Only enters depolarised cells Stains depolarised cells green

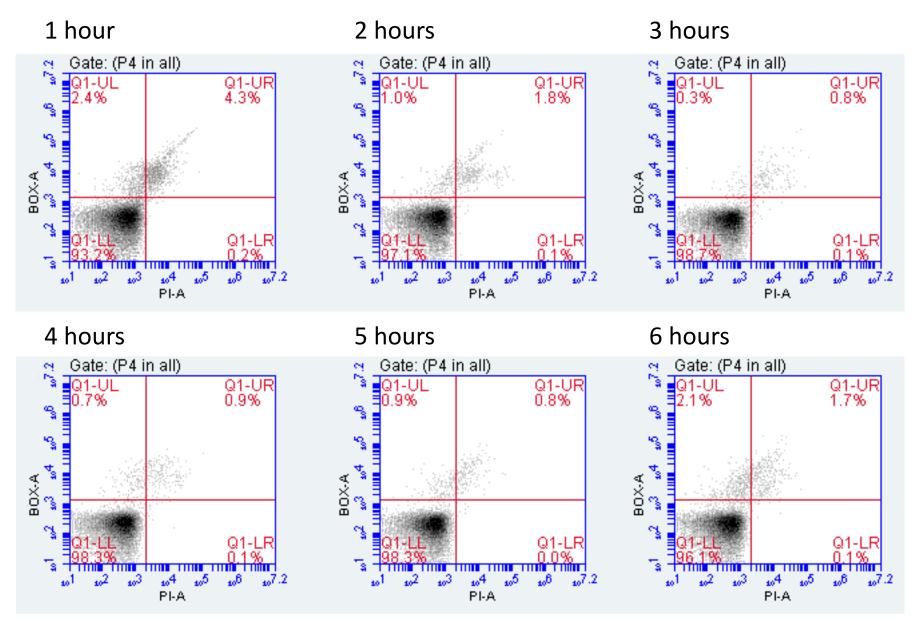
Propidium Iodide (PI)

Enters cells through holes in wall
Stains dead cells red

Advantages:

- Rapid method
- Allows monitoring of VBNC bacteria
- Allows counting of cells

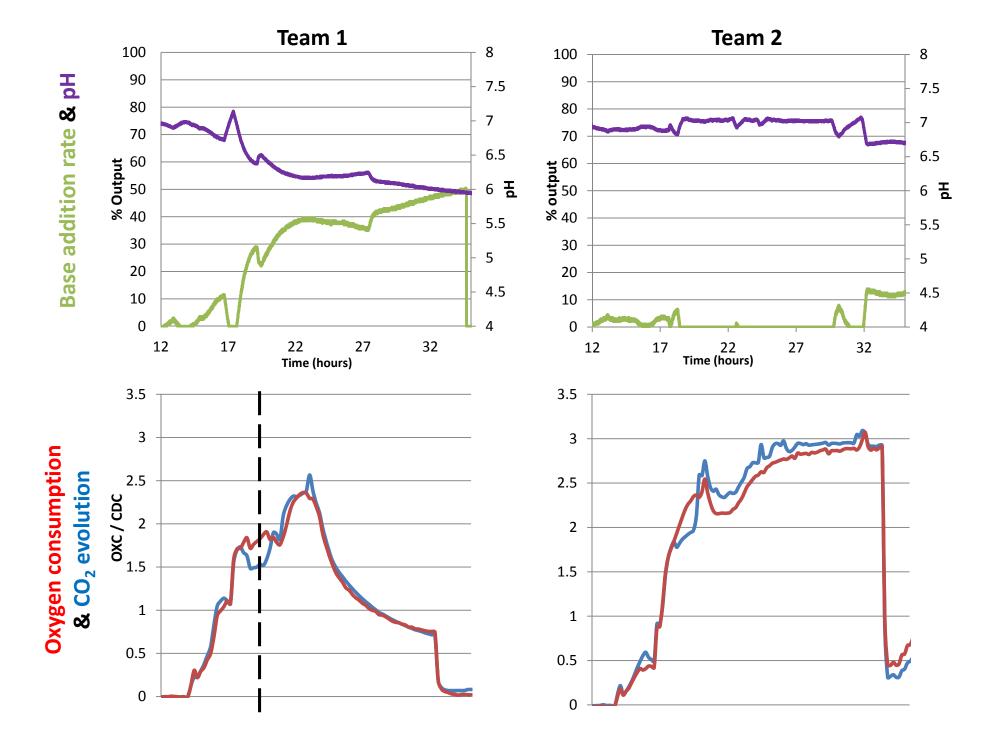
FCM with physiology dyes



Pilot plant

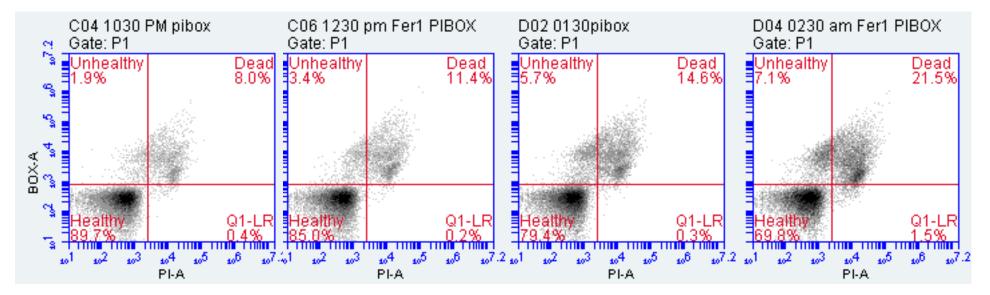
- Growth of *E. coli* in 150 litre vessels over the course of a week
- Fed-batch with glucose
 - Glucose feeding rate is critical to success
- Students encouraged to develop their own analysis methods and strategies – including FCM





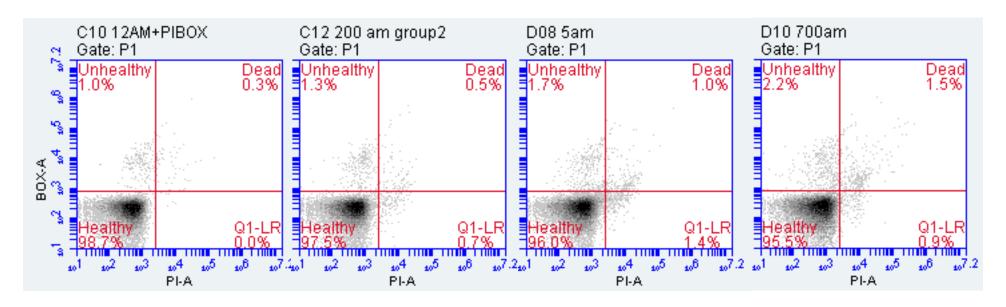
Team 1 – FCM physiology monitoring

More dead and injured cells – reflects poor glucose feed control



Team 2 – FCM physiology monitoring

More healthy cells – good glucose feed control

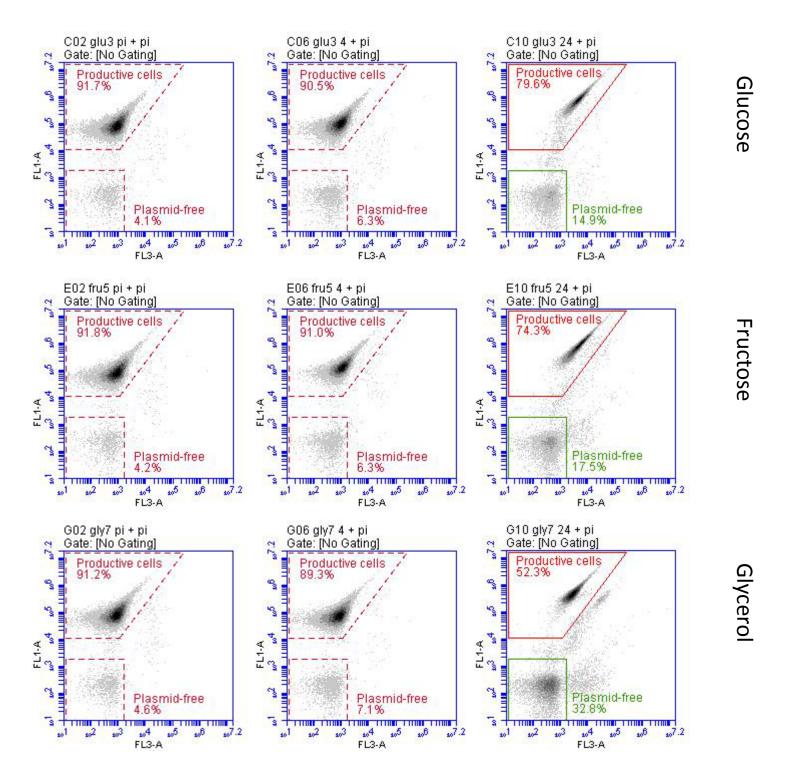


Food microbiology

- Use of flow cytometry as a method for monitoring bacteria during food processing
- Acid resistance and acid adaptation
- Differentiation between bacteria and food matrix

MSc / MEng Research projects

- Monitoring physiology using dyes during biotransformation reactions
- Monitoring poly(3-hydroxybutyrate) (PHB) productivity by bacterial cultures
- Recombinant protein-GFP fusion studies to screen new growth conditions such as choice of carbon source
 - RP-GFP fusions allow measurement of both quantity and folding quality



Doctoral training

- BBSRC-funded MIBTP (Midlands Integrative Biosciences Training Partnership)
 - Warwick, Birmingham & Leicester Universities
 - 52 PhD students per year
- Extensive training programme:
 - Quantitative skills
 - Placements
 - Techniques masterclasses

Doctoral training - master class

- Day 1:
 - Day of lecture / seminar teaching
 - Ability to go into detail and give examples
 - Papers given to students
- Day 2: Reading / preparation
- Day 3:
 - Students give 10 minute talks on papers
 - Practical session

Doctoral training - practical

- *E. coli* wild-type
- E. coli GFP+
- S. cerevisiae
- Comparison of scatter measurements
- Live & Dead cells with a variety of stains (PI, BOX, SYTO9)
- Unknown mixtures (problem-solving aspect)
- Hands-on time is invaluable to apply theory (and test theory)

Challenges and opportunities

- Students have diverse backgrounds
 - Gaps in knowledge (e.g. optics, fluorescence)
 - Students can teach each other
- Students are interested in different areas
 - Comparisons with other techniques
 - Fertile ground for new ideas about methods and measurement techniques
- Students are very keen!

Outlook

- Training in theory of FCM is interdisciplinary and needs life science and physical science knowledge.
- Comparison with previously-used techniques is helpful.
- Combination of taught material with practical experience is very useful.

Acknowledgements

Postdocs: Dr Isaac Vizcaino-Caston & Dr Alfred Fernandez-Castane

PhD students: James Leech, Ikhlaas Kasli, Asma Zulkifly, Hani El Kadri, Hussam Fallatah; Duangkanok Tanangteerapong, Chris Wyre, Amir Anvarian, Louise Hackett

MSc / MEng students: Alifiana Sara, Adna Farah, David Walsh & Christian Mather, Matthew Bridgeman, Mauricio Santos, He Na, Tianqi Wang, Adriana Benarroch, Raúl Mateos González, Ana Álvarez Martín, Shakthiswar Ragu, Kenneth Liu Hung Yaw, Samuel Harvey & David Rothera